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Development of plastic/gelatin bilayer active packaging film with antibacterial and water-absorbing functions for lamb preservation

12

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Abstract

In order to extend the shelf life of refrigerating raw lamb by inhibiting the growth of 13 microorganisms, preventing the oxidation of fat and protein, and absorbing the juice outflow 14 of lamb during storage, an active packaging system based on plastic/gelatin bilayer film with 15 essential oil was developed in this study. Three kinds of petroleum-derived plastic films, 16 oriented polypropylene (OPP), polyethylene terephthalate (PET), and polyethylene (PE), were 17 coated with gelatin to make bilayer films for lamb preservation. The results showed 18 significant improvement in the mechanical properties, oxygen, moisture, and light barriers of 19 the bilayer films compared to the gelatin film. The OPP/gelatin bilayer film was selected for 20 further experiments because of its highest acceptance by panelists. If the amount of juice 21 outflow was less than 350% of the mass of the gelatin layer, it was difficult for the gelatin 22 film to separate from lamb. With the increase in essential oil concentration, the water 23 absorption capacity decreased. The OPP/gelatin bilayer films with 20% mustard or 10% 24 oregano essential oils inhibited the growth of bacteria in lamb and displayed better 25 mechanical properties. Essential oil decreased the brightness and light transmittance of the 26 bilayer films and made the film yellow. In conclusion, our results suggested that the active 27 packaging system based on OPP/gelatin bilayer film was more suitable for raw lamb 28 preservation than single-layer gelatin film or petroleum-derived plastic film, but need further 29 study, including minimizing the amount of essential oil, enhancing the mechanical strength of 30 the gelatin film after water absorption. 31

32 Keywords: active packaging system; chilled lamb; gelatin/plastic bilayer film; water

33 absorption; bacterial inactivation

34 Introduction

Raw meat is a perishable food, however, due to online shopping and globalization of the 35 market, the transportation radius of raw meat has increased, and consumers have higher 36 requirements for the extension of the shelf life of raw meat (Ahmed et al., 2017; Mills & 37 Brightwell, 2014). Microbial spoilage, color deterioration, a decline in eating quality, fluid 38 outflow, lipid oxidation, and inferior types of meat (in particular, dark, firm, and dry and pale, 39 soft, and exudative meats) are the major problems in refrigerating raw lamb meat (Barbosa-40 Pereira et al., 2014; Holman et al., 2018). In addition, these problems lead to food waste, 41 substantial economic losses, and a huge carbon footprint, as discussed in some articles (Rood 42 43 et al., 2022; Shao et al., 2022; Soro et al., 2021). To solve the problems mentioned above, active packaging (AP) has been proposed by meat producers and scientists. AP has extra 44 functions in addition to the characteristics provided by traditional packaging (Ahmed et al., 45 2017; Barbosa-Pereira et al., 2014; Smaoui et al., 2022). Currently, there are mainly two 46 strategies to make AP. The first strategy includes the use of relatively independent accessories 47 with specific functions that exist in the headspace of the packaging. The second strategy to 48 make AP consists of using active substances with functional characteristics in combination 49 with packaging materials (Ghoshal, 2018). 50

Biodegradable materials, especially edible film used as a carrier for different types of active substances, have been increasingly studied in recent years to extend the shelf life of meat and meat products. However, the main goal of these studies was to replace plastic films because of the non-degradable characteristics and problems related to the incineration of petroleumderived plastic polymers commonly used in meat packaging and the migration of toxic and harmful substances such as phthalates into the meat (Harmon & Otter, 2022; Moreira et al., 2015; Soro et al., 2021; Xu et al., 2010). Gelatin is one of the most widely investigated

biopolymers in the field of edible films (Falguera et al., 2011; Gupta et al., 2022; Lacroix & 58 Vu, 2014; Paulo et al., 2021; Sun et al., 2022). It is a protein material derived from the 59 collagen of animal bones or skin after physical or chemical degradation or denaturation. 60 Gelatin can be used as a food additive, an edible packaging film material, and an encapsulant 61 (Etxabide et al., 2016; Moorhouse & Grundon, 1994). It is of great interest in the field of 62 edible films because of its abundant sources (Tongnuanchan et al., 2015), superior film-63 forming properties, better degradability, high biosafety, good nutritional value (de Carvalho & 64 Grosso, 2004), and complete absorption in the body. Edible packaging films made from 65 gelatin have been widely studied for their suitability in various food applications (Nilsuwan et 66 al., 2017). Several of these studies are on the application of monolayer gelatin film in meat 67 and fish products. Some researchers studied the inactivation effect of gelatin-based edible 68 films containing active substances (which were mostly antioxidants and antimicrobials) on 69 spoilage and pathogenic microorganisms of raw meat, including total viable bacterial counts, 70 total mesophilic aerobic bacterial counts, psychrophilic aerobic bacterial counts, Escherichia 71 coli, Bacillus subtilis, Staphylococcus aureus, Listeria monocytogenes, Salmonella 72 Typhimurium, and L. monocytogenes, and obtained good inactivation effects (Albertos et al., 73 2017; Ejaz et al., 2018; Lin et al., 2018; Qiu et al., 2022; Wu et al., 2014; Yilmaz et al., 2022). 74 Some studies investigated the effect of gelatin-based edible films incorporated with active 75 substances on the sensory properties, color, textural properties, total volatile basic nitrogen, 76 thiobarbituric acid reactive substances, and moisture loss of raw meat; and found that these 77 films extended meats' shelf life (Kaewprachu et al., 2015; Qiu et al., 2022; Yilmaz et al., 78 2022). Some researchers found that gelatin-based edible films carrying active substances 79 retarded lipid oxidation and decreased the peroxide values of meat (Hematizad et al., 2021; 80 Kaewprachu et al., 2015; Kaewprachu et al., 2018; Hamann et al., 2022; Syahida et al., 2021). 81 However, few studies have focused on the consumer acceptance of the changes in gelatin 82

83 films during storage and meat quality changes induced by gelatin film packaging during84 storage.

Although gelatin has many advantages, it also has disadvantages when used alone. Raw 85 lamb muscle meat contains more than 70% water (Rao et al., 2018), and during storage, juice 86 outflow of raw lamb meat varies from 0.5% to 15% based on storage conditions, premortem 87 management, and so on (Bond & Warner, 2007; Ekiz et al., 2022; Holman et al., 2020; Moore, 88 1990). After the gelatin film wrapped on the surface of the meat absorbs the juice outflow, the 89 gelatin film disintegrates, and its mechanical strength decreases dramatically. Thus, the use of 90 a monolayer gelatin film in the preservation of fresh meat has many limitations (Ciannamea et 91 al., 2018; Lim et al., 1999; Villasante et al., 2020; Yuan et al., 2021). It is also difficult to 92 develop AP based on petroleum-derived plastic polymers because of the instability of 93 antioxidants and antimicrobials at high temperatures during the production of plastic 94 polymers (Nur Hanani et al., 2018). An approach that combines petroleum-derived plastic 95 polymers with other biopolymers to gain an extra function by mixing antioxidants or 96 antimicrobials has attracted the attention of researchers (Joerger et al., 2009; Nur Hanani et 97 al., 2018). To control the microorganism growth and the sensory quality deterioration of 98 muscles induced by juice outflow, moisture-absorbent sheets, blankets, and pads are usually 99 used for absorbing fluid exudates from raw fresh meat and fish (Ahmed et al., 2017). These 100 existing absorbent sheets on the market can be replaced by gelatin films since they have a 101 strong affinity to moisture and can absorb several times their own weight of water (Kavoosi et 102 al., 2013). 103

Therefore, an AP system based on gelatin/plastic bilayer films with different concentrations of oregano or mustard essential oils was developed in this study to absorb juice outflow and inhibit spoilage microorganisms. In this study, we first determined the maximum weight of lamb that could be wrapped in a given size of gelatin film. Next, the thickness of the gelatin

film was calculated according to the water absorption capacity of the film that could absorb 108 5% juice outflow of raw lamb. Food-grade plastic films of different materials, such as 109 oriented polypropylene (OPP), polyethylene terephthalate (PET), and polyethylene (PE), were 110 used for laminating with gelatin to make bilayer films. The type of plastic film in the bilayer 111 film was selected based on the degree of bonding between the plastic and gelatin layers, the 112 mechanical, oxygen barrier, light barrier properties, and sensory evaluation of the bilayer film 113 by consumers after application to lamb. However, no studies are available on consumers 114 acceptability of the packaging film after water absorption and the sticky phenomenon of the 115 meat surface caused by the gelatin film migrating to the meat surface. Finally, the essential 116 oils were added to the gelatin layer of the bilayer films, and the effects of essential oil 117 concentrations on the mechanical properties, bacterial inactivation, color, and light 118 transmittance were studied to determine the suitable concentration of essential oil. 119

120

121 Materials and methods

122 Preparation of single-layer gelatin film

Gelatin (4 g) was placed in a conical flask containing 300 mL of distilled water and stirred on an intelligent magnetic stirrer (ZNCL-GS, Yike Laboratory Technology Co. Ltd., Guangzhou, Guangdong, China) at a temperature of 60°C and speed of 800 r/min for 1 h. Subsequently, 1.2 g (30% of the weight of the gelatin) of glycerol was added, and the stirring was continued for 0.5 h. Next, the solution was left to cool at lab temperature (21.9°C). Gelatin and glycerol were purchased from Aladdin (Shanghai, China).

The gelatin solution placed at lab temperature $(21.9^{\circ}C)$ in the above step was poured into a 28.5 × 28.5 cm acrylic tank, and then the acrylic tank with gelatin solution was placed in a chamber (JYH-66, Yuejin Medical Equipment Co., Ltd, Shanghai, China) and dried at a temperature of 50°C and relative humidity of 30% for 15 h. After drying, the gelatin film was peeled off after its temperature was cooled to room temperature. Films were placed in a thermostatic incubator at 25°C and 43% relative humidity to further determine the film index.

135 Preparation of OPP/gelatin, PE/gelatin, and PET/gelatin bilayer films

In a 28.5×28.5 cm acrylic tank, the OPP, PE, or PET films were laid flat in the tank, and 136 the same steps were followed as mentioned in the "Preparation of single-layer gelatin film" 137 section. The gelatin solution was poured onto the OPP, PE, or PET film. After drying for 15 h, 138 the OPP/gelatin, PE/gelatin, or PET/gelatin films were taken out and then these double-layer 139 films were placed in a thermostatic incubator at 25°C and 43% relative humidity until the 140 further determination of the film index. PET plastic, PE plastic, and OPP plastic films were 141 purchased from Hongmei paper and plastic packaging factory (Chaozhou, Guangdong, 142 China). 143

144 Preparation of OPP/gelatin bilayer films with essential oil

Gelatin (4 g) was put in a conical flask. Next, 300 mL of distilled water was added to it and 145 stirred with a magnetic stirrer at a temperature of 60°C and a speed of 800 r/min for 1 h. 146 Glycerol (1.2 g) was then added and stirred continuously for 0.5 h. The essential oil (Yuanye, 147 Shanghai, China) and Tween 80 (Aladdin, Shanghai, China) were vortexed at a ratio of 2:1 148 (v/w) for 1 min and mixed thoroughly before adding them to the gelatin solution. The 149 concentrations of oregano essential oil were 0%, 10%, 20%, 30%, and 40%, and those of 150 mustard essential oil were 20%, 40%, 60%, and 80% of the mass of gelatin. The mixed 151 solution was added to the gelatin solution, stirred at room temperature for 1 h, poured into the 152 acrylic tank covered with OPP film, and dried at 50°C in a vacuum drying oven for 15 h. 153

154 Mechanical properties of films

155 The mechanical properties of films were determined based on the ISO 527-1: 2012 method

with some modifications (Jamróz et al., 2021). Film strips $(15 \times 80 \text{ mm})$ were used to measure tensile strength (TS) and elongation at break (EAB) of films, using a texture analyzer (TMS-PILOT, FTC Co., Ltd., Virginia, USA) with an initial separation of 50 mm and a crosshead speed of 200 mm/min. The measurement was made nine times, and the average value was taken. The TS and EAB were calculated using the following equations:

 $TS = Fm/(d \times b)$

- 161
- 162

 $EAB = (L1 - L0)/L0 \times 100\%$

where Fm was the maximum tensile force (N) of the sample; d was the width of the film
(mm); b was the thickness of the film (mm); L0 was the initial length of the sample (mm); and
L1 was the maximum fracture length of the sample (mm).

166 Oxygen resistance properties of films

The oxygen permeability (OP) and oxygen permeability coefficient (OPC) of OPP/gelatin, 167 PE/gelatin, or PET/gelatin bilayer films were determined based on the ISO 2556 method with 168 minor modifications using a gas permeator (VAC-VBS, Languang Electromechanical 169 Technology Co., Ltd., Jinan, Shandong, China). The OP is the volume of oxygen passing 170 through a sample per unit area and per unit time. The OPC is the volume of gas passing 171 through the sample per unit thickness, per unit area, and per unit time at stable transmission. 172 The proportional mode was 10%, the gas replacement time was 60 s, the upper and lower 173 cavity degassing time was 4 h, the temperature was 25°C, relative humidity was 18%, and the 174 gas pressure was 1.01 kgf/cm². 175

176 Water vapor permeability (WVP) of films

The water vapor permeability of films was determined based on the ASTM E96 method with some modifications (Wang et al., 2018; Liang et al., 2018). A permeable cup containing fully dried CaCl₂ particles was sealed with OPP/gelatin, PE/gelatin, or PET/gelatin bilayer 180 films. The permeable cup was then placed in a sealed facility with saturated NaCl solution 181 (osmotic pressure of 1753.55 Pa). The mass of the glass was measured every 1 h until there 182 was no significant change in the mass of the glass. The WVP was calculated as follows: 183 $WVP = (\Delta m \times H)/(\Delta t \times S \times P)$ 184 where Δm was the transit dose of water vapor (g); H was the average thickness of the film 185 (m); S was the test area 0.00196 m²; Δt was the test time (s); and P was osmotic pressure (Pa) 186 of NaCl saturated solution.

187 Light transmittance of films

The light transmittance of films was determined according to Liang and Wang's study (2018). The film strips of 1×4.5 cm size were pressed close to the inner side of the colorimetric dish at lab temperature, and its light transmission in the wavelength range of 200–800 nm was measured. An empty colorimetric dish was used as the control.

192 Color analysis of films

The color of bilayer films with mustard essential oil was measured using a colorimeter (CR-400, Konica Minolta, Co., Ltd., Japan). The whiteboard was used as a reference for color difference measurement. Five locations were selected on each film to measure L^* , a^* , and b^* ; one at the center and four around the perimeter of the film. The color of bilayer films with oregano essential oil was measured using the colorimeter (NR60CP, Sanenchi, Co., Ltd., Shenzhen, Guangdong, China).

199 Water absorption and water dissolution properties of gelatin films

The water absorption and water dissolution properties of the films were expressed in terms of the weight change of gelatin films in water. Films were cut into 100×100 mm sizes and then completely immersed in distilled water. The weight of the films was measured at a specific time after being immersed in water. The weight change ratio was calculated using the following formula: Weight change ratio = (the weight of film soaked in water for a certain period – the weight of film before being soaked in water)/the weight of film before being soaked in water.

The water dissolution properties of gelatin films were also determined by the protein migration from the film to water when the films were immersed in water. Four portions of the same mass of dried films were put into a conical flask, 50 mL of distilled water with a temperature of 4°C was added and placed in a 4°C refrigerator, and the protein content of the water was quantified using the BCA protein assay (Thermo Fisher Scientific Inc., Waltham, USA) at 1, 2, 3, and 4 d.

Adhesiveness of gelatin film after water absorption

A double compression test was performed to evaluate the adhesiveness of gelatin film after 214 water absorption. The films with or without essential oil were cut into sizes of 10×10 cm. Ten 215 layers were stacked together, placed into distilled water, soaked for a certain period, and taken 216 out, and the double compression test was measured using a texture analyzer (TMS-PILOT, 217 FTC Co., Ltd., Virginia, USA) according to the method described in a previous study 218 (Fiszman et al., 2000). The parameter settings used were as follows: TA5 probe model, an 219 initial force of 0.15 N, test rate of 60 mm/min, test distance of 20 mm, 50% deformation, and 220 interval time of 5 s. Adhesiveness is the negative force during the first bite, representing the 221 force necessary to pull the compressing plunger away from the sample. 222

223 Sensory evaluation of bilayer films and gelatin film after packing lamb

The *longissimus dorsi* muscles from both sides of ten small tail Han sheep raised in the same pen and of 7 months of age were selected and slaughtered in Jinhongqingzhen Co. Ltd., Baoding, Hebei, China. After slaughtering, the muscles were taken from both sides of the carcass and frozen in liquid nitrogen. After complete freezing, the meat was stored in a

refrigerator at -80°C, and before use, it was thawed to 4°C for storage experiments. Lamb 228 was divided into meat pieces of about 400 g, wrapped with monolayer gelatin film, 229 OPP/gelatin bilayer film, PE/gelatin bilayer film, or PET/gelatin bilayer film. Packaged 230 muscles stored at 4°C for 5 days were evaluated by ten trained panelists according to the 231 sensory evaluation criteria (Table 1). Ten pieces of meat were wrapped in each packaging 232 material. When one panelist arrived to assess the film and the lamb, 4 out of the 10 pieces of 233 meat in each packaging material were randomly selected for evaluation. After evaluation, the 234 meat was re-wrapped for the next panelist's assessment test. The higher the score, the higher 235 the acceptability of the film. All procedures described here were undertaken following the 236 principles and guidelines of the Ethics Committee of Hebei Agricultural University, and the 237 approval number is 2021095. 238

Antibacterial properties of bilayer films 239

Lamb meat was soaked in saline, and the lamb was slapped to transfer the bacteria to the 240 saline. The saline containing lamb bacteria was diluted to 1×10⁵ CFU/mL, and 1 mL was 241 added to the Petri dish. The plate counting agar was poured and shaken up. After the saline 242 was dried, a 4×4 cm double-layer film containing essential oil was placed in the center of the 243 Petri dish. The film without essential oil was used as a blank control. The Petri dishes were 244 cultured at 37°C for 48 h, and the number of colonies in the plate was counted. Bacterial 245 counts were converted to CFU/g before statistical analysis. 246

Antibacterial properties of gelatin/OPP essential oil film used in vacuum 247

packaging system 248

The preparation of OPP/gelatin essential oil film used in the vacuum packaging system was 249 carried out according to the method described in the previous section "Preparation of 250 OPP/gelatin bilayer films with essential oil" with some modifications. In this experiment, the 251

essential oil was a mixture of mustard and oregano essential oils. The ratio of mustard and 252 oregano essential oils was 1:4, and the total amount of essential oil was 20% of the weight of 253 gelatin (4 g). Lamb longissimus dorsi muscles were divided into meat pieces of about 400 g, 254 packaged in vacuum bags (control group) or vacuum bags + oregano-mustard composite 255 essential oil double layer film (OPP/gelatin essential oil film as the inner film wrapped on the 256 surface of lamb, and then put into the vacuum plastic bag) (treatment group), and stored at 257 4 °C. Three packages of these lambs from each treatment group were taken out every 3 days 258 and the total colony count in the samples was measured. The total colony count of bacteria 259 was detemined as follows: 1 g meat was placed in 9 mL saline. The bacteria of the meat was 260 transferred to the saline by beating. One mL of saline was injected into a petri dish. Then 261 AGAR medium was injected to the petri dish. Medium and bacterial saline was mixed and 262 cooled in lab temperature. After the medium has solidified, petri dish were cultured for 2 days 263 at 37°C. The number of colonies in the plate was counted. Bacterial counts were converted to 264 CFU/g before statistical analysis. 265

266 Data analysis

Data were analyzed using SPSS software v.22.0 (IBM Corporation Inc., USA), and results were expressed as mean \pm standard deviation. One-way analysis of variance was used to compare data differences among multiple groups. Duncan's test and LSD method were used for multiple comparisons of mean values. P < 0.05 indicated a statistically significant difference. The graphs were created using the Origin 2019 software (OriginLab, Northampton, MA, USA).

273

274 Results and Discussion

275 Mechanical properties of films

The TS indicates the ability of the material to resist stretching and is a key indicator for 276 evaluating the mechanical properties of the film. As seen in Fig. 1, the order of TSs from 277 largest to smallest was as follows: OPP/gelatin film > PET/gelatin film > PE/gelatin film > 278 gelatin film. The TS of single-layer gelatin film was 2.80 MPa; meanwhile, the TSs of bilayer 279 OPP/gelatin, PE/gelatin, and PET/gelatin films increased by 8.58, 2.94, and 5.71 times, 280 respectively, compared to the gelatin film. As observed for the gelatin film in this work, the 281 TSs of some kinds of gelatin in other studies were also lower than 3.5 MPa, which could be 282 due to the different sources or extraction methods of gelatin, the processing technology of the 283 film, and the addition of plasticizer and other substances (Haghighi et al., 2019; Rather et al., 284 2022; Said & Sarbon, 2022). However, as a food packaging material, sufficient tensile 285 strength is generally needed; the greater the TS of the film, the better the food is protected. In 286 some countries, the TS of the food packaging film must be greater than 3.5 MPa (Kim et al., 287 1995). Thus, compared to the single-layer gelatin film, the plastic film present in the bilayer 288 film improved the TS of the bilayer film. 289

EAB is the deformation of the film before the film breaks during the stretching process, and films with excessive deformation are not suitable for application in packaging. As seen from the data in Fig. 1, the EAB of single-layer gelatin film was 190%, while those of OPP/gelatin, PE/gelatin, and PET/gelatin films decreased by 145%, 175%, and 147%, respectively, compared with the gelatin film.

In summary, the gelatin film laminated with the plastic film was more suitable for application in the packaging of raw meat products. From the point of view of mechanical strength, compared with the other three kinds of films, OPP/gelatin bilayer film was the best.

298 Oxygen resistance of films

The lower the OP and OPC of the film, the stronger its ability to restrict oxygen entry in the packaging of raw meat, which can reduce the oxidation of meat products and prolong their

storage period. As shown in Fig. 2, PET/gelatin and OPP/gelatin bilayer film had better 301 oxygen resistance than PE/gelatin bilayer film. The oxygen permeability and oxygen 302 permeability coefficient of the single-layer gelatin film was not shown in Fig. 2, as the OP of 303 the film was too large for the gas permeator to measure. This indicated that the oxygen 304 permeability of single-layer gelatin film was bigger than that of the plastic/gelatin bilayer 305 films. Many studies have determined that the gelatin film has a good oxygen resistance 306 capacity (Ciannamea et al., 2018; Etxabide et al., 2022). According to the previous studies, 307 the OPC of gelatin film was between 0.01-68492 cm³·µm·m⁻²·d⁻¹·kPa⁻¹ (Hosseini et al., 308 2016; Lee & Song, 2017; Nur Hanani et al., 2013; Tyuftin & Kerry, 2021; Wang et al., 2009). 309 This was higher than the OPC of OPP/gelatin, PE/gelatin, and PET/gelatin bilayer films, 310 which showed a value between 0.004–0.0026 cm³· μ m·m⁻²·d⁻¹·kPa⁻¹. 311

The difference in OPC can be induced by factors such as differences in film thickness, pH, 312 relative humidity, gelatin source, plasticizer concentration (Ciannamea et al., 2018). The 313 approach used in this study could allow bubble formation in the gelatin film and introduce 314 variation in film thickness; the higher the film thickness, the bigger the OPC (Tyuftin & 315 Kerry, 2021). The OP of gelatin film is also affected by the glycerol concentration; the higher 316 the glycerol concentration, the higher the OP values (Lim et al., 1999). The OP of gelatin 317 films is also influenced by pH; the higher the pH, the lower the oxygen permeability of the 318 gelatin film (Nur Hanani et al., 2014). 319

320 Moisture resistance of films

The water vapor transmission rate of a film reflects the water barrier effect of the film (Rhim et al., 2000; Du et al., 2011) and should be low enough to avoid moisture loss on the surface of the meat. As shown in Fig. 3, the water vapor transmission coefficient of gelatin film was 5.1×10^{-8} g/(s·m·Pa). However, the water vapor transmission coefficients of

OPP/gelatin, PET/gelatin, and PE/gelatin bilayer films were significantly lower than that of 325 the gelatin film, and the values were 0.47×10^{-8} g/(s·m·Pa), 0.81×10^{-8} g/(s·m·Pa), and 326 0.39×10^{-8} g/(s·m·Pa), respectively. The larger the transmission coefficient, the worse the 327 moisture barrier performance; therefore, compared to OPP, PET, and PE films, gelatin film 328 had poorer moisture barrier performance, which was consistent with the results of other 329 studies (Etxabide et al., 2022; Tyuftin & Kerry, 2021; Villasante et al., 2020). In addition, 330 there was no significant difference in moisture barrier performance among the three bilayer 331 films. 332

Light resistance of films

The light resistance performance is an important indicator of the film, and the transmittance 334 can indirectly represent the magnitude of the film's light resistance. As shown in Fig. 4 (a), all 335 the films had a strong ultraviolet absorption at 200-280 nm, which was consistent with the 336 results obtained by other researchers (Nilsuwan et al., 2018; Wang et al., 2018). The gelatin 337 film had good resistance to ultraviolet radiation at 200-280 nm, due to the rich content of 338 tyrosine and tryptophan that absorbed ultraviolet radiation in protein films such as gelatin 339 (Ejaz et al., 2018). Ultraviolet light can cause changes in nutrients and flavor substances of 340 341 raw meat and accelerate lipid oxidation, inducing its deterioration. Therefore, the more ultraviolet light absorption of the film, the lower the transmittance, the slower the lipid 342 oxidation rate of food, and the better the quality of raw meat during storage. Fig. 4 (a) shows 343 that the bilayer film improved the light resistance of gelatin film, which could be better for 344 maintaining the stability of the food system. A wavelength of 700 nm was selected to analyze 345 the differences between the four films at visible light transmittance. As shown in Fig. 4 (b), 346 compared with the monolayer gelatin film, the light resistance of the double-layer films 347 improved to a greater extent. However, all the values of the double-layer films were between 348

349 88% and 91%, and no significant differences were observed between the three bilayer films.

350 Sensory evaluation of films and lamb after 5 days storage at 4°C

From the sensory evaluation results in Fig. 5, it can be seen that no drying occurred on the 351 surface of the lamb meat in the bilayer packaging, but the surface of the lamb packed with a 352 monolayer gelatin film had an obvious drying phenomenon. The surface of lamb meat 353 wrapped in four kinds of films was somewhat sticky because the ingredients in gelatin 354 migrated to the meat surface, which was acceptable to panelists; the score of these films was 355 above 6 points. Some juice outflow was seen in the package when it was wrapped with either 356 a bilayer film or a monolayer gelatin film. This might be because the lamb used in this study 357 exhibited a higher purge loss during storage, indicating the need for a thicker gelatin film. 358

Due to the water-absorbing property, the monolayer gelatin film was more severely cracked and due to the drying of the surface of the lamb wrapped in a monolayer gelatin film, the acceptability of this film was the lowest among others. The score for the acceptability of the monolayer gelatin film was only 2 points, suggesting that the gelatin film was highly unacceptable to panelists. The overall acceptability of the samples wrapped in OPP/gelatin bilayer film was higher as its flexibility was the highest of all bilayer films.

365 Water absorption and water dissolution properties of films

As seen in Table 2, the weight change rate of the gelatin layer with different concentrations of mustard or oregano essential oils was 3-5 when films were immersed in 4°C water for 1 day. This implied that after being wrapped in OPP/gelatin bilayer film, the gelatin film absorbed the water of the lamb before the juice outflow occurred. A slight reduction in the weight of meat is usually observed in the early stages of storage (Kaewprachu et al., 2018). However, this study showed that the loss of meat storage in the middle and late stages of storage was more likely due to the loss of water through the gelatin film into the air, inducing

the drying of the meat surface. As shown in Fig. 5, the acceptance of the lamb decreased when 373 the surface of the lamb was dry. Some evaluators also indicated that the dried meat surface 374 was unacceptable, thus limiting the application of monolayer pure gelatin film in meat 375 storage. In some studies, there was less storage loss of meat wrapped in gelatin film, which 376 might be due to the differences in the methods used to measure the storage loss of meat. These 377 studies wrapped the meat in gelatin film, placed them in a plastic bag, and then measured the 378 weight loss during storage. The gelatin film could absorb the moisture released by the meat, 379 thus lowering the storage loss (Kaewprachu et al., 2015). 380

The value of the water absorption rate of the gelatin film was close to 550%, as the gelatin 381 film could absorb more than 5 times its mass of water because of the relatively hydrophilic 382 nature of gelatin (Etxabide et al., 2022; Kaewprachu et al., 2015; Yang et al., 2022). With a 383 juice loss rate of 5% for fresh lamb, the thickness of the film could be calculated by the mass 384 of fresh lamb that needed to be wrapped by the gelatin film and the water absorption rate of 385 gelatin. With the increase in the amount of essential oil added, the mass change rate of the 386 gelatin film gradually decreased. Moreover, the water absorption of the film decreased 387 because the tight film structure could be destroyed by essential oil molecules, making the film 388 easier to dissolve. Meanwhile, Ciannamea et al. (2018) found that the hydrophobic character 389 of essential oil droplets within the matrix could obstruct the migration of water through the 390 film. 391

When the gelatin film was immersed in 4°C water for more than 2 days, the weight of the film decreased, which was consistent with the findings of a previous study (Ahammed et al., 2020). This was mainly due to glycerin (Liu et al., 2017) and protein in the film dissolving into the water (Table 2). Mustard and oregano essential oil significantly increased the solubility of gelatin films, which was consistent with the results of a previous study (Kavoosi et al., 2013). The dissolution rate of the gelatin film decreased from 1 d to 4 days. It was speculated that the faster growth rate on 1 and 2 days was because most of the soluble proteinwas dissolved; therefore, the growth rate dropped on 3 days.

At the later stage of lamb storage, the gelatin film still maintained its film shape and 400 attached to the OPP plastic film after absorbing the juice out of the raw lamb, making this 401 edible film extremely easy to be peeled off from the raw lamb and not dissolve totally into the 402 raw lamb. When the OPP/gelatin bilayer film was incorporated with essential oil, it 403 disintegrated after some days of packaging the lamb (data not shown). Therefore, increasing 404 the intactness of gelatin films containing essential oil after absorbing water will be a hot 405 research topic for the future development of gelatin AP. To date, few studies are available 406 related to this topic. Some scholars have confirmed that tannic acid addition could keep the 407 films prepared using a mixture of dialdehyde glucomannan and gelatin intact after being 408 immersed in water (Yuan et al., 2021). It should be noted that the focus is not on enhancing 409 410 the hydrophobic characteristics of the gelatin film with essential oil but on improving the intactness, flexibility, and mechanical strength of the gelatin film after water absorption. The 411 results of studies related to enhancing the mechanical properties of gelatin hydrogels or 412 sponges might provide some references that could be useful for strengthening the mechanical 413 strength of gelatin films after absorbing water in this experiment. For example, the Maillard 414 reaction between polysaccharides and gelatin was a useful method to strengthen the links 415 between the molecules of gelatin sponges; however, whether the intactness of gelatin film was 416 maintained after being immersed in water needs further study (Yang et al., 2022). Gelatin 417 modifications, pH adjustment, temperature control, and ion addition might be other effective 418 methods to enhance the strength of gelatin gel (Bao et al., 2019; Farris et al., 2009; Huang et 419 al., 2019) after absorbing water. 420

421 Adhesiveness of gelatin films with essential oil after water absorption

422 Adhesiveness is the energy or a force required to overcome the attraction between the food

surface and the surface contact with other substances. As seen in Table 3, the adhesiveness of 423 gelatin film was 1.9 N in 1 min after water absorption, then the adhesiveness started to 424 decrease, and it dropped to 0.15 N after 5 min. Thus, the thickness of the gelatin film should 425 be determined considering both the water absorption and adhesiveness characteristics of the 426 gelatin film. If adhesiveness is very large, the gelatin film can adhere to the meat when 427 unpacking, and the film and meat may not be easy to separate. When the time was between 1– 428 3 min, the water absorption rate of gelatin was close to 4 times the gelatin mass. Therefore, 429 the weight of the gelatin film needs to be less than a quarter of the juice outflow to lower the 430 adhesiveness of the gelatin film. For the gelatin film with essential oil, the maximum 431 adhesiveness of the gelatin film decreased with the addition of essential oil. 432

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434 Effect of essential oil on mechanical and antibacterial properties of OPP/gelatin

435 bilayer film

As shown in Table 4, when the concentration of essential oil increased, the TS first 436 increased and then decreased. This increase might be attributed to an appropriate amount of 437 essential oil embedded in gelatin molecules, improved interaction between molecules, and 438 increased TS. As the concentration continued to increase, the gelatin molecules could not 439 encapsulate more essential oil molecules, and the agglomeration of the essential oil molecules 440 resulted in the formation of yellow bumps on the surface of the films. An excess of essential 441 oil formed a discontinuous and non-uniform film structure, thus weakening the network 442 structure of the film and reducing the TS of the film. With the increase in essential oil, the 443 EAB of the film decreased, which might be because the addition of essential oil increased the 444 interaction force between gelatin molecules, increased the strength of the film, and decreased 445 the EAB. However, some researchers have found different results. Martucci et al. (2015) 446 found a decrease in TS and a stable value of EAB in gelatin films incorporated with oregano 447

essential oil. Carvacrol is one of the main components of oregano essential oil. Kavoosi et al.
(2013) found a decrease in TS and an increase in EAB in gelatin films containing carvacrol,
glycerol and glutaraldehyde. The difference between the results in this study and those in the
published articles might be caused by the different concentrations of essential oil used in
gelatin film. The concentrations of essential oils used in this study were several to ten times
that of the concentrations reported in the literature.

The antibacterial properties of OPP/gelatin bilayer films with different essential oil 454 concentrations are shown in Table 4. The present study showed that gelatin films with 455 essential oils could inhibit the growth of bacteria in lamb. Mustard and oregano essential oils 456 were effective at concentrations of more than 10% and 20%, respectively, which was similar 457 to the concentration of antimicrobial effectiveness of protein films containing essential oils 458 (Martucci et al., 2015). But, colony count was not much reduced at high concentration of 459 essential oils group. We have reason to doubt whether this concentration could extend the 460 shelf life of meat. The possible reasons were as follows. The antibacterial activity of plant 461 essential oil depends on its active components. The main components of oregano essential oil 462 are carvacrol and thymol, while that of mustard essential oil is allyl isothiocyanate (Jayasena 463 & Jo, 2013). In the preparation of the bilayer film, the emulsified essential oil was added, and 464 films were dried at 50°C, which could cause the volatilization of allyl isothiocyanate, 465 carvacrol, and thymol. Meanwhile, allyl isothiocyanate could react with the free amino acids 466 in the gelatin. Allyl isothiocyanate also degraded during the mixing and drying of the film. 467 When drying was complete, these effects led to a decrease in mustard and oregano essential 468 oils in the bilayer films. The remaining essential oil after volatilization combined with the 469 components of the gelatin film, which was another reason for the decreased antimicrobial 470 effectiveness of essential oil, preventing the migration of essential oil to agar (Burt, 2004; 471 Javasena & Jo, 2013; Martucci et al., 2015). The gelatin composition and the emulsification 472

process might also be related to the migration rate of essential oil from gelatin film to agar orfood surface.

In this study, the bacterial inactivation of oregano essential oil was stronger than that of 475 mustard essential oil. Moreover, mustard oil is more flavorful than oregano oil, which could 476 cause significant changes in food senses and non-acceptance by consumers. The combination 477 of oregano and mustard essential oils can be an excellent way to add them to the AP system. 478 The changes in the total colony count of refrigerated lamb packaged in a vacuum bag + 479 OPP/gelatin film containing a combination of oregano-mustard essential oil are shown in Fig. 480 6. The results showed that the OPP/gelatin film with essential oils prolonged the shelf life of 481 vacuum-packaged lamb. Since essential oil migrating from edible films such as gelatin to 482 food is a slow process, the concentration of essential oil on the surface of food can be 483 maintained within a certain range for a long time, thus extending the shelf life of food. In this 484 way, it is more effective to inhibit the growth of microorganisms than to spray essential oil 485 directly on the surface of products (Quintavalla & Vicini, 2002). 486

487 Effect of essential oil on color and light transmittance of OPP/gelatin bilayer

488 film

As reported in other articles, gelatin film exhibited good transparency (Table 5) (Etxabide 489 et al., 2022). The light transmittance of the films reduced with increased essential oil 490 concentration, which could be attributed to the astigmatism of essential oil, increasing the 491 light resistance of the film (Ejaz et al., 2018). The more the addition of essential oil, the 492 stronger the astigmatism effect and the lower the light resistance of the film. As shown in 493 Table 6, the light transmittance at 700 nm was significantly influenced by the essential oil 494 concentration. The more the essential oil concentration, the lower the transmittance of the film 495 at 700 nm. 496

The film's color was affected by the natural color of essential oils (Said & Sarbon, 2022). 497 As seen in Table 6, after adding the oregano or mustard essential oils, the brightness of the 498 gelatin film kept decreasing and the color became increasingly darker. Furthermore, the film 499 tended to turn yellow, and the b^* value became significantly bigger. The consumers prefer the 500 color of the lamb to be red. They do not consider the lamb fresh when it is yellow in color, 501 especially in China. The yellow color decreases consumers' willingness to select meat at the 502 point of purchase (Tomasevic et al., 2021). Therefore, the concentrations of oregano and 503 mustard essential oils used in gelatin films should be less than 20% and 10%, respectively. 504

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507 Conclusion and Prospect

508 After comprehensively considering the mechanical properties, oxygen barrier, light barrier, and moisture barrier and sensory evaluation of OPP/gelatin, PET/gelatin, and PE/gelatin 509 510 bilayer films, OPP was selected as the binding layer to the gelatin surface. Compared with food-grade plastic films, the OPP/gelatin bilayer film with mustard or oregano essential oils 511 could absorb the juice outflow of raw lamb and inhibit the growth of the bacteria of lamb. 512 Compared with monolayer gelatin film, the mechanical properties, oxygen barrier, moisture 513 barrier, and light barrier properties and consumer acceptance of the OPP/gelatin bilayer film 514 were improved. Thus, OPP/gelatin bilayer film incorporated with 10% oregano essential oil 515 was more suitable for application to the packaging of raw lamb meat. Bilayer films improved 516 the degree of gelatin fracture after water absorption, but increasing the intactness of gelatin 517 films containing oregano essential oil after absorbing water, and reducing the loss of essential 518 oils during the drying of the bilayer film are still urgent research topics for the future 519 development of gelatin AP. 520

522 Conflicts of Interest

523 The authors declare no potential conflicts of interest.

524

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532

533 Author Contributions

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540 Ethics Approval

541 This article were undertaken following the principles and guidelines of the Ethics Committee

of Hebei Agricultural University, and the approval number is 2021095.

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Table 1. Sensory evaluation criteria of bilayer films and single layer gelatin films after packing lamb

	<u> </u>	0	1 0	1 0
	8-10 points	6-7 points	4-5 points	1-3 points
Drying of lamb surface	No drying	Slight drying	Some drying	Severe drying
Slipperiness of lamb surface	Not slippery	Slight slippery	Some slippery	Severe slippery
Degree of film damage	No damage	Slight damage	Some damage	Severe damage
Juices running out of lamb	No juices	Slight juices	Some juices	Severe juices
Flexibility of film	Fully acceptable	Acceptable	Unacceptable	Highly unacceptable
Overall acceptability (lamb and film)	Fully acceptable	Acceptable	Unacceptable	Highly unacceptable
()				

Table 2. Weight change ratio and protein solubility of single layer gelatin films with different
 concentrations of mustard or oregano essential oils at 4°C in water.

/ <u>24 conc</u>	entration								Oregan	gano Essential Oil			
	Time	0%	20%	40%	60%	80%	-	0%	10%	20%	30%	40%	
	1 day	5.50± 0.31ª	4.48±0. 55 ^b	4.01±0. 63 ^{bc}	3.26±0. 26 ^{cd}	3.11±0. 28 ^d		5.50±0. 32ª	4.21±0. 59 ^b	3.50±0. 83 ^{bc}	3.11± 0.40°	2.95±0 .24°	
Weigh change	2 day	$\begin{array}{c} 5.40 \pm \\ 0.21^{a} \end{array}$	4.39±0. 38 ^b	4.00±0. 20 ^b	3.22±0. 58°	3.05±0. 41°		5.40±0. 21ª	4.10±0. 16 ^{ab}	2.96±0. 26 ^b	3.13± 0.21 ^b	2.86±0 .11 ^b	
ratio	3 day	$\begin{array}{c} 5.39 \pm \\ 0.45^a \end{array}$	4.32±0. 35 ^b	3.96±0. 31 ^b	3.23±0. 34°	3.12±0. 62°		5.39±0. 45ª	4.10±0. 80 ^b	2.90±0. 24°	3.10± 0.14°	2.80±0 .45°	
	4 day	$\begin{array}{c} 5.23 \pm \\ 0.23^{a} \end{array}$	4.12±0. 23 ^b	3.83±0. 35 ^b	3.12±0. 20°	3.10±0. 42°		5.23±0. 23ª	3.91±0. 12 ^{ab}	2.69±0. 63 ^b	$2.85 \pm 0.29^{\rm b}$	2.71±0 .13 ^b	
	Time		0.	.539						0.550			
P-value	Essential Oil 0		000				0.000						
	Time×	Essential	Oil 1	.000						1.000			
	1 day	0.34 ± 0.01^{dD}	2.14±0. 01 ^{cC}	3.02 ± 0.02^{bC}	3.23±0. 03 ^{aC}	3.39 ± 0.06^{aC}		0.34±0. 01 ^{eD}	1.24±0. 02 ^{dD}	1.98±0. 04 ^{cD}	$\begin{array}{c} 3.02 \pm \\ 0.16^{\mathrm{bB}} \end{array}$	$3.23{\pm}0$.04 ^{aC}	
Protein solubility	2 day	$\begin{array}{c} 0.41 \pm \\ 0.01^{\text{dC}} \end{array}$	2.56±0. 07 ^{cB}	3.23±0. 09 ^{bB}	3.31 ± 0.08^{abC}	3.43 ± 0.06^{aC}		0.41±0. 01 ^{dC}	1.64±0. 06° ^C	2.44±0. 08 ^{ьв}	$\begin{array}{c} 3.23 \pm \\ 0.09^{\mathrm{aA}} \end{array}$	3.31±0 .08 ^{aC}	
(mg/mL)	3 day	$\begin{array}{c} 0.43 \pm \\ 0.08^{eB} \end{array}$	2.84±0. 09 ^{dA}	3.31±0. 09° ^B	3.43±0. 06 ^{bB}	3.57±0. 02 ^{aB}		0.43±0. 08 ^{eB}	1.91±0. 04 ^{dB}	2.33±0. 02 ^{cC}	$\begin{array}{c} 3.31 \pm \\ 0.18^{\text{bA}} \end{array}$	$\begin{array}{c} 3.43{\pm}0\\.06^{aB}\end{array}$	
	4 day	$\begin{array}{c} 0.46 \pm \\ 0.01^{\text{dA}} \end{array}$	2.97±0. 08° ^A	3.55±0. 08 ^{bA}	3.66±0. 07 ^{abA}	$3.72\pm0.08^{\mathrm{aA}}$		0.46±0. 01 ^{eA}	2.04 ± 0.03^{dA}	2.69±0. 05 ^{cA}	$\begin{array}{c} 3.33 \pm \\ 0.09^{bA} \end{array}$	3.60±0 .06 ^{aA}	
	Time		0.	.000						0.000			
P-value	Essentia	al Oil	0.	.000			0.000						
	Time×	Essential	Oil 0	.000						0.000			

For each essential oil, lowercase letters in each row represent significant differences (P < 0.05). For each

essential oil, capital letters in each column represent significant differences (P < 0.05).

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731	Table 3. Adhesiveness of gelatin films with 10% oregano or 20% mustard essential oils after
732	absorbing water for different times at 4°C.

Time	Gelatin film	Gelatin film with 10% oregano oil	Gelatin film with 20% mustard oil
1 min	-1.91 ± 0.30^{bC}	-0.35 ± 0.09^{aC}	-0.32 ± 0.04^{aB}
2 min	-1.32 ± 0.28^{bB}	-0.27 ± 0.05^{aB}	-1.35 ± 0.13^{bC}
3 min	-0.40 ± 0.16^{A}	-0.30 ± 0.00^{BC}	-0.30 ± 0.05^{B}
4 min	-0.20 ± 0.04^{aA}	-0.27 ± 0.05^{bB}	-0.30 ± 0.02^{bB}
5 min	-0.17 ± 0.04^{A}	-0.15 ± 0.02^{A}	-0.15 ± 0.00^{A}
10 min	$-0.15 \pm 0.00^{\text{A}}$	-0.15 ± 0.01^{A}	-0.15 ± 0.01^{A}
60 min	-0.15 ± 0.01^{A}	-0.15 ± 0.00^{A}	$-0.15 \pm 0.00^{\text{A}}$
3 days	$-0.15 \pm 0.00^{\text{A}}$	-0.15 ± 0.00^{A}	-0.15±0.00 ^A
	Time	0	.000
P-value	Film	0	.000
	Time × Film	0.	.000

733 Lowercase letters in each row represent significant differences (P < 0.05). Capital letters in each column

734 represent significant differences (P < 0.05).

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739 Table 4. Mechanical and antibacterial properties of OPP/gelatin bilayer films with different

Essential Oil	Concentration (%)			Colony count (log CFU/mL)
	0	26.81±0.52 °	158.35±28.84 ª	$6.48{\pm}0.01$ ^a
	10	61.23±5.22 ª	60.66±2.63 ^b	6.31±0.02 ^b
Oregano Essential Oil	20	43.20±2.31 ^b	37.39±6.32 ^b	$6.17{\pm}0.05$ °
	30	31.54±2.45 °	30.28 ± 7.08 ^b	$6.14{\pm}0.07$ °
	40	28.79±3.69 °	29.37±5.08 ^b	$6.05 {\pm} 0.08$ °
	0	26.81±0.52 ^b	158.35±28.84 ª	6.48±0.01 ^a
	20	29.81±3.37 ^b	56.36±12.33 ^b	6.35±0.03 ^b
Mustard Essential Oil	40	32.12±2.53 ^b	36.33±6.98 bc	$6.29{\pm}0.05$ ^b
	60	74.92±8.21 ª	32.47 ± 8.85 bc	$6.14{\pm}0.07$ °
	80	38.02±5.10 ^b	15.85±4.77 °	6.02 ± 0.03 ^d

740 concentrations of oregano or mustard essential oils

For each essential oil, lowercase letters in each column represent significant differences (P < 0.05).

742 Antibacterial properties was done as follow: bilayer film containing essential oil was placed in the center of

the Petri dish containing bacteria. The stronger the inhibition of bacteria, the fewer colonies could grow in

the Petri dish.

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Wavelength	Mustard Essential Oil						Oregano Essential Oil			
(nm)	0%	20%	40%	60%	80%	0%	10%	20%	30%	40%
200	1.93±0. 12 ^{bF}	1.57±0. 21 ^{cD}	2.17±0.0 5 ^{bE}	2.63±0. 12 ^{aD}	2.04±0. 13 ^{bE}	3.63±0. 31 ^{bE}	3.67±0. 05 ^{bC}	3.40±0. 08 ^{bD}	4.37±0.0 9 ^{aD}	4.23±0 05 ^{aF}
300	66.23 ± 2.85^{aD}	36.43± 3.53 ^{bC}	14.63±2. 60 ^{cD}	12.73± 2.42 ^{cC}	10.23± 0.76 ^{cD}	$\begin{array}{c} 66.23 \pm \\ 2.85^{aD} \end{array}$	$\begin{array}{c} 63.33 \pm \\ 4.86^{aB} \end{array}$	50.07 ± 4.13^{bC}	47.53±7. 69 ^{bcC}	38.43 2.73 ^{cl}
400	$\begin{array}{c} 86.53 \pm \\ 0.54^{aC} \end{array}$	77.87± 1.39 ^{bB}	68.00±1. 96° ^C	66.13± 3.97 ^{cdB}	$\begin{array}{c} 62.77 \pm \\ 1.28^{\text{dC}} \end{array}$	$\begin{array}{c} 86.53 \pm \\ 0.54^{aC} \end{array}$	85.77 ± 2.35^{aA}	$75.40\pm 2.05^{\mathrm{bB}}$	71.03±1. 98 ^{bB}	66.60 0.86 ^{cl}
500	88.63± 1.91 ^{aBC}	83.27 ± 1.62^{aA}	74.97±3. 24 ^{bB}	73.67 ± 2.84^{bA}	73.20± 4.12 ^{ьв}	88.63± 1.91 ^{aBC}	88.33 ± 1.76^{aA}	81.30± 1.24 ^{bA}	76.63±1. 16 ^{cAB}	73.17 0.71 ^d
600	$\begin{array}{c} 89.30 \pm \\ 2.33^{aBC} \end{array}$	$83.67 \pm 1.48^{\mathrm{bA}}$	77.73±1. 80° ^{AB}	75.30± 1.43° ^A	74.50± 2.35 ^{cB}	$\begin{array}{c} 89.30 \pm \\ 2.33^{aBC} \end{array}$	88.03 ± 2.55^{aA}	82.31 ± 0.94^{bA}	79.30±1. 88 ^{bcA}	76.37 1.09 ^{cE}
700	$\begin{array}{c}92.03\pm\\1.32^{aAB}\end{array}$	$83.63 \pm 2.80^{\mathrm{bA}}$	79.47±1. 32 ^{bcAB}	75.23± 3.77 ^{cA}	76.97± 2.15 ^{cAB}	92.03± 1.32 ^{aAB}	91.27 ± 0.17^{aA}	83.57 ± 0.82^{bA}	80.53±1. 35°A	78.71 1.64⊄
800	94.33± 3.22ªA	84.97± 2.27 ^{bA}	80.80±1. 48 ^{bA}	80.67± 5.07 ^{bA}	$80.17 \pm 1.70^{\mathrm{bA}}$	94.33 ± 3.22^{aA}	91.30± 0.22ªA	84.13± 1.02 ^{bA}	82.00±1. 42 ^{bA}	80.07 1.87 ^b
	Wavelen	gth		0.000				0.000		
P-value	Essential	l Oil		0.000				0.000		
	Wavelen	gth ×Essei	ntial Oil	0.000				0.000		

Table 5. Light transmittance of OPP/gelatin bilayer films with different concentrations of mustard essential oil or oregano essential oil at 4°C.

748 For each essential oil, lowercase letters in each row represent significant differences (P < 0.05). For each

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Table 6. Color difference and light transmittance of OPP/gelatin bilayer films with different
 concentrations of oregano or mustard essential oils at 700 nm.

Essential Oil	Concentration (%)			CIE b*	Transmittance at 700 nm (%)
	0	55.42±0.80 ª	2.81±0.27 ^a	3.36±0.34 °	92.03±1.32 ª
Oregano	10	53.43±0.53 ^b	2.31±0.14 ^b	4.29±0.24 ab	91.27±0.17 ^a
Essential	20	51.23±0.55 °	1.82±0.22 °	3.94±0.12 ^b	83.57±0.82 ^b
Oil	30	51.81 ± 1.15 ^{cd}	1.79±0.19 °	3.90±0.50 ^b	80.53±1.35 °
	40	50.79±0.93 ^d	2.14±0.16 ^b	4.56±0.16 ^a	78.70±1.64 °
	0	67.81±0.29 ^a	11.01±0.39 ^a	9.76±0.63 ^d	92.03±1.32 ^a
Mustard	20	63.38±0.86 ^b	8.70±1.21 ^b	9.93±0.68 ^{cd}	83.63±2.80 ^b
Essential Oil	40	59.66±1.11 °	8.54±1.03 ^b	10.67±0.5 bc	79.47±1.32 bc
	60	58.79±1.31 ^{cd}	8.13±0.66 ^b	10.83±0.45 ^b	75.23±3.77 °

57.2±1.53 d 80 6.79±0.57 ° 13.18±0.38 a 76.97±2.15 °

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OPP/gelatin PE/gelatin PET/gelatin

For each essential oil, lowercase letters in each column represent significant differences (P < 0.05). 754 755 756 757 758 Tensile strength Elongation at break 30 а A 200 25 Elongation at break (%)

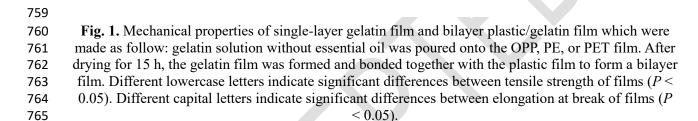
Tensile strength (MPa)

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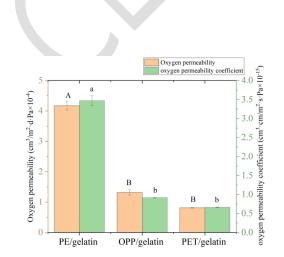
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773 Fig. 2. Oxygen permeability and oxygen permeability coefficient of plastic/gelatin bilayer films which were made as follow: gelatin solution without essential oil was poured onto the OPP, PE, or PET film. 774 After drying for 15 h, the gelatin film was formed and bonded together with the plastic film to form a 775 bilayer film. Different lowercase letters indicate significant differences between oxygen permeability 776 coefficient of films (P < 0.05). Different capital letters indicate significant differences between oxygen 777 permeability of films (P < 0.05). 778

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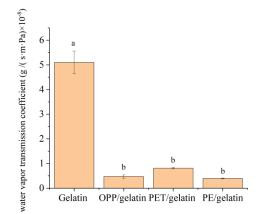


Fig. 3. Water vapor transmission coefficient of single-layer gelatin film and bilayer plastic/gelatin film which were made as follow: gelatin solution without essential oil was poured onto the OPP, PE, or PET film. After drying for 15 h, the gelatin film was formed and bonded together with the plastic film to form a bilayer film. Different lowercase letters indicate significant differences between the water vapor transmission coefficient of films (P < 0.05).

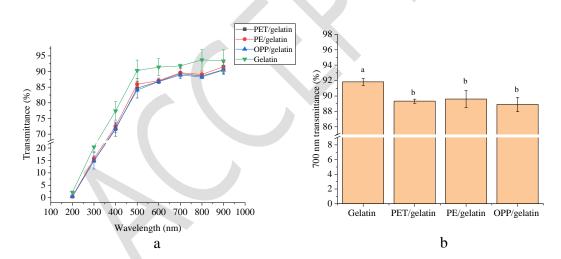


Fig. 4. Transmittance from 100–1000 nm (a) and transmittance at 700 nm (b) of gelatin, PET/gelatin,
 PE/gelatin, and OPP/gelatin bilayer films which were made as follow: gelatin solution without
 essential oil was poured onto the OPP, PE, or PET film. After drying for 15 h, the gelatin film was
 formed and bonded together with the plastic film to form a bilayer film. Different lowercase letters in
 (b) indicated significant differences between films (P < 0.05)

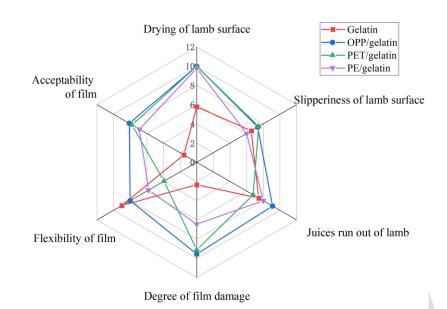
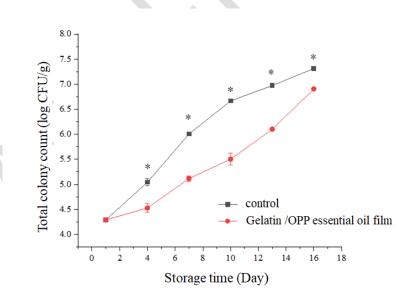


Fig. 5. Sensory evaluation of single layer gelatin film, bilayer films, and lamb wrapped in different
 films. Bilayer plastic/gelatin films were made as follow: gelatin solution without essential oil was
 poured onto the OPP, PE, or PET film. After drying for 15 h, the gelatin film was formed and bonded
 together with the plastic film to form a bilayer film

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Fig. 6. Changes in the total colony count of refrigerated lamb (4°C) packaged in vacuum bag or vacuum bag + OPP/gelatin film containing a combination of essential oils. The compound ratio of mustard and oregano essential oil was 1:4, and the total amount of essential oil was 20% of the quantity of gelatin. Treatment group: Packaging with vacuum bag + oregano-mustard composite essential oil in double layer film (OPP/gelatin film with essential oils as the inner film wrapped on the surface of lamb, and then put into the vacuum plastic bag). Control group: Lamb packaged in vacuum bag. Asterisk indicate significant differences between control and treatment group (P < 0.05).