

TITLE PAGE
- Food Science of Animal Resources -
Upload this completed form to website with submission

ARTICLE INFORMATION	Fill in information in each box below
Article Type	Short communication
Article Title	Effect of <i>Lactiplantibacillus plantarum</i> on Osteoporosis in the Ovariectomized Rat
Running Title (within 10 words)	Effect of <i>L. plantarum</i> on Osteoporosis
Author	Eun-Sun Jin ^{1,2} , Ji Yeon Kim ² , JoongKee Min ² , Sang Ryong Jeon ^{2,3} , Kyoung Hyo Choi ^{2,4} , Shehzad Abid Khan ⁶ , Gi-Seong Moon ^{5,6*} and Je Hoon Jeong ^{2,7*}
Affiliation	¹ Department of Internal Medicine, College of Medicine, Kyung Hee University ² Laboratory of Stem Cell Therapy, College of Medicine, Asan Medical Center, University of Ulsan ³ Department of Neurological surgery, Asan Medical Center, University of Ulsan, College of Medicine ⁴ Department of Rehabilitation Medicine, Asan Medical Center, University of Ulsan, College of Medicine ⁵ Department of Biotechnology, Korea National University of Transportation ⁶ 4D Convergence Technology Institute, Korea National University of Transportation, Jeungpyeong 27909, Korea ⁷ Department of Neurosurgery, Soonchunhyang University Bucheon Hospital
Special remarks – if authors have additional information to inform the editorial office	Je Hoon Jeong and Gi-Seong Moon contributed equally to this work and should be considered co-corresponding authors.
ORCID (All authors must have ORCID) https://orcid.org	Eun-Sun Jin(https://orcid.org/0000-0003-1182-8244) Ji Yeon Kim(https://orcid.org/0000-0002-5869-9649) JoongKee Min(https://orcid.org/0000-0002-1680-4797) Sang Ryong Jeon(https://orcid.org/0000-0002-8340-7978) Kyoung Hyo Choi(https://orcid.org/0000-0001-9137-3889) Shehzad Abid Khan(https://orcid.org/0000-0002-2814-5573) Gi-Seong Moon(https://orcid.org/0000-0003-3033-5250) Je Hoon Jeong(https://orcid.org/0000-0002-4656-0113)
Conflicts of interest List any present or potential conflict s of interest for all authors. (This field may be published.)	The authors declare no potential conflict of interest.
Acknowledgements State funding sources (grants, funding sources, equipment, and supplies). Include name and number of grant if available. (This field may be published.)	This work was supported by the National Research Foundation of Korea (NRF) grant funded by the Korea government (MSIT) (grant no. NRF-2017R1A2B1005327), and this work was supported by the Soonchunhyang University Research Fund. This research was also supported by Basic Science Research Program through the National Research Foundation of Korea(NRF) funded by the Ministry of Education(No. 2021R1A6A1A03046418).
Author contributions (This field may be published.)	Conceptualization, E.S.J., G.S.M., and J.H.J.; Investigation, J.Y.K.; Methodology, J.M. and G.S.M.; Resources, J.M. and G.S.M.; Visualization, S.R.J. and K.H.C.; Writing - original draft, E.S.J., S.A.K., G.S.M., and J.H.J.; Writing - review & editing, E.S.J., J.Y.K., J.M., S.R.J., K.H.C., S.A.K., G.S.M., and J.H.J. All authors have read and agreed to the published version of the manuscript.
Ethics approval (IRB/IACUC) (This field may be published.)	This experiment was approved by the Institutional Animal Care and Use Committee of Asan Institute for Life Sciences, Seoul, Republic of Korea (2017-14-228).

CORRESPONDING AUTHOR CONTACT INFORMATION

For the <u>corresponding</u> author (responsible for correspondence, proofreading, and reprints)	Fill in information in each box below
First name, middle initial, last name	Gi-Seong Moon; Je Hoon Jeong

Email address – this is where your proofs will be sent	gsmoon@ut.ac.kr; neuri71@schmc.ac.kr
Secondary Email address	
Postal address	Major of Biotechnology, Korea National University of Transportation, 61 Daehak-ro, Jeungpyeong 27909, Korea; Department of Neurosurgery, Soonchunhyang University Bucheon Hospital, Soonchunhyang University College of Medicine, 170, Jomaru-ro, Bucheon 14584, Korea
Cell phone number	+82-10-2388-3460; +82-10-8147-6132
Office phone number	+82-43-820-5251; +82-32-621-6709
Fax number	+82-43-820-5272; +82-32-621-5016

7

8

Preliminary Study on Effect of *Lactiplantibacillus plantarum* on Osteoporosis in the Ovariectomized Rat

Abstract

Osteoporosis is a growing global health concern primarily associated with decreased estrogen in postmenopausal women. Recently, some strains of probiotics were examined for potential anti-osteoporotic effects. This study intended to evaluate the impacts of *Lactiplantibacillus plantarum* MGE 3038 strain (MGE 3038) in ovariectomized rats. For this purpose, twelve weeks old female Wistar rats (n=21; 250–300 g) were divided into 3 groups; ovariectomy (OVX) group, OVX/MGE 3038 group and Sham group (control). In these groups; two went through respective OVX and one had daily MGE 3038 administration through oral gavage. Prior to 16 weeks after OVX, we collected blood samples and extracted the tibiae. We scanned the extracted tibiae by in-vivo micro-computed tomography (micro-CT) and evaluated pathology by hematoxylin and eosin (H&E) and Masson's trichrome staining. The serum levels of C-telopeptide of type I collagen (CTX), osteocalcin (OC), and the receptor activator of nuclear factor- κ B ligand (RANKL) were examined. The OVX/MGE 3038 group showed increases in bone mineral density, trabecular bone volume, trabecular number, and trabecular thickness (Tb.Th), and a decrease in trabecular spacing than the OVX group. However, OVX/MGE 3038 group and control group were measurably comparable in Tb.Th. Micro-CT, H&E, and Masson's trichrome findings exhibited increased preservation and maintenance of trabecular bone structure in the OVX/MGE 3038 group in comparison to the OVX group. In serum, the levels of CTX, OC and RANKL were significantly different between the OVX and OVX/MGE 3038 groups ($p<0.05$). Taken together, *L. plantarum* MGE 3038 could be helpful for the treatment of osteoporosis.

Keywords: *Lactiplantibacillus plantarum*; probiotics; osteoporosis; ovariectomy; animal model

Introduction

As populations have aged, osteoporosis has become a significant public health issue around the world. Osteoporosis is related with an imbalance in metabolism of bone whereby bone resorption is more than bone formation (Christiansen, 1992). Some treatments, including hormone replacement therapy, bisphosphonates, vitamin D and calcium supplementation, selective estrogen-receptor modulators, and teriparatide® have been displayed to effectively decrease the risk of bone fracture and loss in patients of osteoporotic (Collins et al., 2017; Kanis et al., 2013). Non-pharmacological therapies include a nutritional diet, exercise, and surgical treatment for fractures (Collins et al., 2017; Kanis et al., 2013). Some previous studies have reported potential benefits of probiotic supplementation on bone disease and some strains of probiotics have been shown to be effective treatments for osteoporosis in experimental animal models (Collins et al., 2017). The motivation behind this study was to evaluated the effects of *Lactiplantibacillus plantarum* MGE 3038, which was previously isolated from a natural cheese and might influence on bone health with other cheese components, supplementation on ovariectomized rats.

Materials and Methods

Animals

Twelve weeks old female Wistar rats (n=21; 250–300 g) were divided into 3 groups; ovariectomy (OVX) group, OVX and *L. plantarum* MGE 3038 (OVX/MGE 3038) group and

Sham group (control). In these three groups; two went through respective OVX and one had daily MGE 3038 administration through oral gavage. This trial was permitted by the Animal Care and Use Committee of Asan Institute for Life Sciences, Seoul, Republic of Korea (2017-14-228).

Ovariectomy

Ovariectomy was achieved through a small skin incision in the back area under 5% isoflurane. We performed a peritoneal dissection to reach the ovary before cutting the uterine horn and Fallopian tube after vessel ligation. Finally, we removed the ovaries. After wound closure, an intramuscular antibiotic (cefazolin, 10 mg/kg) was injected for three days.

Probiotic treatment

The MGE 3038 used in this study was previously isolated from a natural cheese and its partial 16S rRNA gene sequence was deposited in GenBank database (accession number OP269658) and has been stored at -80°C prior to use. Four weeks after OVX surgery, probiotic supplementation began. Over sixteen weeks, OVX/MGE 3038 group rats were supplied once daily 400 µL of MGE 3038 treatment containing ca. 10⁸ CFU. The OVX and Sham groups were administered with same volume of sterile water.

Micro-computed tomography (CT) evaluation

We evaluated in-vivo CT (Skyscan NV, Kontich, Belgium) at 0, 5, 10, 15 weeks after OVX under the inhalation anesthesia. Finally we extracted both tibiae and evaluated by high-resolution micro-CT (Skyscan NV) 16 weeks after OVX. The volume of interest was confined

between a starting point 2.5 mm distal from the proximal growth plate of the tibia to the diaphysis; the volume was divided over 100 slices. An area of special interest was demarcated in the compartment of whole trabecular bone. The exposure time was 670 ms. Skyscan software (Skyscan NV) was used for image reconstruction and analysis. The trabecular bone volume (BV/TV; %), trabecular thickness (Tb.Th; mm), bone mineral density (BMD; mg/cm³), trabecular number (Tb.N; /mm), and trabecular separation (Tb.Sp; mm) were evaluated.

Pathological evaluation

For the assessment of hematoxylin- and eosin- staining (H&E), all pathological specimens were first decalcified. Afterward, specimens were cut in 3 μ m thicknesses longitudinally. The samples were then deparaffinized and dehydrated. The sections were stained for 5 min with Hematoxylin-I (YD Diagnostics, Yongin, Korea) and Eosin Y (Sigma-Aldrich, USA) for 2 min 30 s. For the Masson's trichrome stain, we treated samples with a mordant solution (5 % iron alum) for 30 min in a 56°C dry oven. Following this, the sections were stained for 10 and 5 min using Weigert's iron hematoxylin nuclear staining (Sigma-Aldrich) and Biebrich scarlet-acid fuchsin (Sigma-Aldrich) solutions, respectively. Then, above stained sections were decolorize by treating for 5 min in a phosphotungstic-phosphomolybdic acid solution. Finally, samples were placed in solution of 1% glacial acetic acid for 3 min.

Blood biochemical assay

To collect the blood from heart, the heart was punctured, enough blood was collected and subjected to centrifugation for serum separation at 1,100 g for 10 min at 4 °C. Subsequently,

type I collagen C-telopeptide (CTX), osteocalcin (OC), and the receptor activator of nuclear factor- κ B ligand (RANKL) was examined using a Rat CTX ELISA Kit (Cusabio, USA), Rat-Mid OC ELISA kit (IDS, UK), and Rat RANK ELISA kit (Cusabio), respectively. The serum parameters were evaluated using a reader of microplate absorbance (Sunrise; TECAN, Switzerland).

Statistical analysis

All analyses were achieved using R (version 3.3.2; The R Foundation for Statistical Computing, Vienna, Austria). Two-way ANOVA with Bonferroni post-test correction and Mann-Whitney U test for body weight and micro-CT measurements were conducted to define whether the parameters differed significantly. All data are presented as means \pm standard deviations (SD) or means \pm standard error (SEM). *p*-value of 0.05 or less was considered statistically significant.

Results

Body weight

After ovariectomy, it was found that the mean body weight increased over the twenty-weeks in all tested groups. A significant increase in the weight of OVX and OVX/MGE 3038 treated groups were observed when compared with the control group. However, the mean body weight of OVX group was higher than the OVX/MGE 3038 group and the difference between the two groups increased over time (data not shown).

Micro-CT measurements

In micro-CT images, changes in osteoporosis were observed in OVX and OVX/MGE 3038 group. The preservation of trabecular structure was more prominent in the OVX/MGE 3038 than in OVX (Fig. 1). The maximum values of BMD, BV/TV, Tb.N, and Tb.Th, and the lowest value of Tb.Sp were observed in control group, while the Tb.Th value difference among the control group and OVX/MGE 3038 group was not significant. However, increased values in BMD, BV/TV, Tb.N, and Tb.Th, and a decrease in Tb.Sp were noticed in the OVX/MGE 3038 group as compared with the OVX group (data not shown).

Pathological evaluation

In H&E and Masson's trichrome-stained sections, decreased bony trabeculae and increased adipose tissue infiltration of the bone marrow were exhibited in the OVX and OVX/MGE 3038 groups compared to control group. The maintenance of trabecular bone structure in the OVX/MGE 3038 group was improved in comparison with the OVX group (Fig. 2).

Blood biochemical assay

Serum levels of CTX and OC among the control group and OVX group were significantly different ($p < 0.05$). However, no meaningful differences were observed in between the control group and OVX/MGE 3038 group ($p > 0.05$). While, among the OVX and OVX/MGE 3038 group, serum levels of CTX and OC were also different significantly ($p < 0.05$) (Fig. 3). Additionally the receptor activator of nuclear factor- κ B ligand (RANKL) was the highest in the OVX group, with significant difference between OVX (113.54 ± 37.99 , ng/mL) and control group (80.821 ± 11.64). However, there was no difference between control and OVX/MGE 3038 group (92.63 ± 11.11) ($p > 0.05$).

Discussion

Probiotics known as good or beneficial live microorganisms, have proved to have positive effects on host health when administered in a sufficient amount (Seddik et al., 2017). Among the lactic acid bacterial species, *L. plantarum* is a well-known probiotic bacterium that can be largely found in fermented food and nutritional products (Seddik et al., 2017), vegetables (Cherdyntseva et al., 2016), beef (Schillinger and Lücke, 1989), herb (Seddik et al., 2017), wine (Berbegal et al., 2016), as well as the gastrointestinal, urogenital, and vaginal tracts (Al Kassaa et al., 2014; Jose et al., 2015). The impressive ability of this bacterium to exist across a large range of environments derives from a high degree of metabolic pathway diversities (Fiocco et al., 2010). The MG 3038 used in this study was previously isolated from a natural cheese.

For postmenopausal osteoporosis examination, a number of studies have been conducted using different animal models (Kalu, 1991; Turner, 2001). Among these animal models, the OVX rat model is mostly used to study postmenopausal osteoporosis, as this rat model is easy to handle and cheaper (Jin et al., 2019). This model shows cancellous bone change including postmenopausal bone loss (Kalu, 1991; Turner, 2001) and is also used as an obesity model (Jeong et al., 2015). The mean body weight in all groups after ovariectomy increased over the twenty-weeks. However, the body weight of rats in the OVX group was higher than the rats of OVX/MGE 3038 group (data not shown). In the micro-CT findings, it was observed that the trabecular structure of rats in the OVX/MGE 3038 group is well maintained in comparison with OVX group (Fig. 1). The trabecular formation was also better maintained in the rats of OVX/MGE 3038 group than in the rats of OVX group as shown in the pathological findings (Fig. 2). These data allow us to support the hypothesis that MGE 3038 is effective for

preventing osteoporotic changes in ovariectomized rats.

Osteoporosis is caused by an age-related decrease of osteoblast progenitor cells in the bone marrow and reduction in estrogen levels as well (Friedenstein, 1976). Some probiotics have shown the potential for bone structure protection in osteoporosis by reducing aging-induced bone loss in senescence-enhanced mice (Kimoto-Nira et al., 2007) while others enhanced bone thickness (McCabe et al., 2013) or improved mineral contents in the cortical bone of chicken (Mutuş et al., 2006). While, the principle mechanism and underlying effects of MGE 3038's treatment have not been fully clarified and are still being investigated. Some hypotheses have proposed that enhanced antimicrobial peptide secretion, host immune system regulation by altering the intestinal microflora, or increased mucus production to regulate luminal pH in the gut thereby improving the absorption of calcium may be responsible for the effects of MGE 3038 supplementation on osteoporotic bone. Kim et al. (2019) presented that *L. plantarum* NK3 and *Bifidobacterium longum* NK39 might alleviate osteoporosis in a ovariectomized mouse model by controlling NF- κ -B-linked TNF- α expression via gut microbiota modulation. In another previous study, *Lactobacillus paracasei* strain alone or mixed with *L. plantarum* strains showed protection effects for ovariectomy-induced cortical bone loss and resorption via regulation of inflammatory cytokines and inhibitor of osteoclastogenesis and influenced on frequency of regulatory T cells in bone marrow in the ovariectomized mouse model (Ohlsson et al., 2014). Recently gut microbiome has been highlighted since it is systematically linked to human physiology such as brain function, immune system, skin health, etc. In particular, osteoporosis could be influenced by gut microbes which modulates nutrient absorption, immune cell balance, and neurotransmitters via gut-brain axis (Ding et al., 2020). Even though more sophisticated researches on the relationship between gut microbiome and bone metabolism should be done under interdisciplinary approach which includes experts including

microbiologist, immunologist, bone pathologist, etc., gut microbiome diversity seems to be one of important factors for bone homeostasis. Alteration of diversity and composition of gut microbiome influences on gut barrier permeability and metabolites, which can trigger inflammatory reaction of specific immune cells including T_H17 cells associated with bone degradation via stimulating the differentiation of osteoclasts in bone marrow (Sato et al., 2006; Cooney et al., 2021). Probiotics including *L. plantarum* are well known to present positive effects on gut health via modulation of gut microbiome, increase of short-chain fatty acids (SCFAs), improvement of gut barrier integrity, etc., which could be closely related to bone health (Hemarajata and Versalovic, 2013).

In our study, after ovariectomy in rats, we evaluated BMD and other parameters for 20 weeks through micro-CT. The OVX/MGE 3038 group displayed an increased BMD, Tb.Th, Tb.N, and BV/TV, and a reduction in Tb.Sp in comparison with the rats of OVX group, although no significant differences were observed statistically. Furthermore, in the case of Tb.Th values no significant differences were noticed between the control group and OVX/MGE 3038 group (data not shown).

Bone remodels to new bone usually achieve by replacement of damaged and old tissue. Osteoclasts and osteoblasts co-work simultaneously in the remodeling unit. Bone remodeling is most protrusive on surface of the cancellous bone, where osteoblast induces new bone formation with the removal of the old and damaged tissues from the eroded surface through osteoclasts (Abdul-Majeed et al., 2012). During the bone resorption stage, serum CTX, making a good bone resorption marker that generally well-reflects osteoclast activity, is increased by osteoclasts. In this study, the CTX levels in the OVX group showed the maximum level and in comparison to the control group displayed significant difference statistically. However, the CTX levels in the OVX/MGE 3038 group were not significantly different from the control

group and have significant difference from the OVX group ($p<0.05$; Fig. 3). During the bone formation process, OC levels in the serum increases (Ivaska et al., 2004; Romero Barco et al., 2012; Rosen et al., 2000). Osteoblasts and osteocytes secrete serum OC that serve as a marker for the bone formation (Szulc and Delmas, 2008). In present study, the serum level of OC in the rats of OVX group was highest among the three groups and significant difference was observed in comparison with the rats in the control and OVX/MGE 3038 groups ($p<0.05$; Fig. 3). Furthermore, RANKL was the highest in the OVX group, with significant difference between OVX and control group. However, there was no difference between control and OVX/MGE 3038 group ($p>0.05$). So, we thought that the LAB strain can improve the osteoporosis via prevention of the activation of the osteoclasts. Taken together, OVX-induced osteoporotic rats showed strong activation of osteoclasts and coupling of osteoblast activation, in which MGE 3038 administration showed preventive effects. Therefore, MGE 3038 may prove beneficial effect on osteoporosis prevention.

As with any study, there are limitations to take into consideration. We did not evaluate changes in the colon, which should include measures of short-chain fatty acids, luminal pH, and mineral absorption (Moslehi-Jenabian et al., 2010; Parvaneh et al., 2014).

Conclusion

L. plantarum MGE 3038 is thought to be helpful in preventing and treating osteoporosis in ovariectomized animal models. Nevertheless the results are preliminary and have limitations to fully understand the mechanisms underlying the effects. In the future, we will identify the active components from the probiotic combined with gut microbiome composition and analyze more sophisticated biomarkers including ratios of osteoblast vs. osteoclast.

Conflicts of Interest

The authors declare no potential conflict of interest.

Acknowledgements

This work was supported by the National Research Foundation of Korea (NRF) grant funded by the Korea government (MSIT) (grant no. NRF-2017R1A2B1005327) and by the Soonchunhyang University Research Fund. This research was also supported by Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Education (No. 2021R1A6A1A03046418).

References

- Abdul-Majeed S, Mohamed N, Soelaiman IN. 2012. Effects of tocotrienol and lovastatin combination on osteoblast and osteoclast activity in estrogen-deficient osteoporosis. *Evid Based Complement Alternat Med* 2012:960742.
- Al Kassaa I, Hamze M, Hober D, Chihib NE, Drider D. 2014. Identification of vaginal lactobacilli with potential probiotic properties isolated from women in north lebanon. *Microb Ecol* 67:722-734.
- Berbegal C, Peña N, Russo P, Grieco F, Pardo I, Ferrer S, Spano G, Capozzi V. 2016. Technological properties of *Lactobacillus plantarum* strains isolated from grape must fermentation. *Food Microbiol* 57:187-194.
- Cherdyntseva TA, Kotova IB, Netrusov AI. 2016. The isolation, identification and analyses of lactobacillus genus bacteria with probiotic potential. *Adv Exp Med Biol* 897:103-111.
- Christiansen C. 1992. Prevention and treatment of osteoporosis: A review of current modalities. *Bone* 13 Suppl 1:S35-39.

267 Collins FL, Rios-Arce ND, Schepper JD, Parameswaran N, McCabe LR. 2017. The potential
268 of probiotics as a therapy for osteoporosis. *Microbiol Spectr* 5 (in press).

269 Cooney OD, Nagareddy PR, Murphy AJ, Lee MKS. 2021. Healthy gut, healthy bones:
270 Targeting the gut microbiome to promote bone health. *Front Endocrinol* 11:620466.

271 Ding K, Hua F, Ding W. 2020. Gut microbiome and osteoporosis. *Aging Dis.* 11:438–447.

272 Fiocco D, Capozzi V, Collins M, Gallone A, Hols P, Guzzo J, Weidmann S, Rieu A, Msadek T,
273 Spano G. 2010. Characterization of the ctsr stress response regulon in *Lactobacillus*
274 *plantarum*. *J Bacteriol* 192:896-900.

275 Friedenstein AJ. 1976. Precursor cells of mechanocytes. *Int Rev Cytol* 47:327-359.

276 Hemarajata P, Versalovic J. 2013. Effects of probiotics on gut microbiota: mechanisms of
277 intestinal immunomodulation and neuromodulation. *Therap Adv Gastroenterol* 6:39-51.

278 Ivaska KK, Hentunen TA, Vääräniemi J, Ylipahkala H, Pettersson K, Väänänen HK. 2004.
279 Release of intact and fragmented osteocalcin molecules from bone matrix during bone
280 resorption in vitro. *J Biol Chem* 279:18361-18369.

281 Jeong JH, Park J, Jin ES, Min J, Jeon SR, Kim DK, Choi KH. 2015. Adipose tissue-derived
282 stem cells in the ovariectomy-induced postmenopausal osteoporosis rat model. *Tissue*
283 *Eng Regen Med* 12:28-36.

284 Jin ES, Kim JY, Min JK, Jeon SR, Choi KH, Lee MS, Jeong JH. 2019. Bilateral ovario-
285 hysterectomy induced osteoporotic rabbit model. *J Biol Regul Homeost Agents* 33:391-
286 396.

287 Jose NM, Bunt CR, Hussain MA. 2015. Comparison of microbiological and probiotic
288 characteristics of lactobacilli isolates from dairy food products and animal rumen
289 contents. *Microorganisms* 3:198-212.

290 Kalu DN. 1991. The ovariectomized rat model of postmenopausal bone loss. *Bone Miner*

15:175-191.

Kanis JA, Mccloskey EV, Johansson H, Cooper C, Rizzoli R, Reginster JY. 2013. European guidance for the diagnosis and management of osteoporosis in postmenopausal women. Osteoporos Int 24:23-57.

Kim DE, Kim JK, Han SK, Jang SE, Han MJ, Kim DH. 2019. *Lactobacillus plantarum* NK3 and *Bifidobacterium longum* NK49 alleviate bacterial vaginosis and osteoporosis in mice by suppressing NF- κ B-linked TNF- α expression. J Med Food 22:1022-1031.

Kimoto-Nira H, Suzuki C, Kobayashi M, Sasaki K, Kurisaki J, Mizumachi K. 2007. Anti-ageing effect of a lactococcal strain: Analysis using senescence-accelerated mice. Br J Nutr 98:1178-1186.

Mccabe LR, Irwin R, Schaefer L, Britton RA. 2013. Probiotic use decreases intestinal inflammation and increases bone density in healthy male but not female mice. J Cell Physiol 228:1793-1798.

Moslehi-Jenabian S, Pedersen LL, Jespersen L. 2010. Beneficial effects of probiotic and food borne yeasts on human health. Nutrients 2:449-473.

Mutuş R, Kocabagli N, Alp M, Acar N, Eren M, Gezen SS. 2006. The effect of dietary probiotic supplementation on tibial bone characteristics and strength in broilers. Poult Sci 85:1621-1625.

Ohlsson C, Engdahl C, Fåk F, Andersson A, Windahl SH, Farman HH, Movérare-Skrtic S, Islander U, Sjögren K. 2014. Probiotics protect mice from ovariectomy-induced cortical bone loss. PLoS One 9:e92368.

Parvaneh K, Jamaluddin R, Karimi G, Erfani R. 2014. Effect of probiotics supplementation on bone mineral content and bone mass density. ScientificWorldJournal 2014:595962.

Romero Barco CM, Manrique Arijia S, Rodríguez Pérez M. 2012. Biochemical markers in

osteoporosis: Usefulness in clinical practice. *Reumatol Clin* 8:149-152.

Rosen HN, Moses AC, Garber J, Iloputaife ID, Ross DS, Lee SL, Greenspan SL. 2000. Serum
 ctx: A new marker of bone resorption that shows treatment effect more often than other
 markers because of low coefficient of variability and large changes with
 bisphosphonate therapy. *Calcif Tissue Int* 66:100-103.

Sato K, Suematsu A, Okamoto K, Yamaguchi A, Morishita Y, Kadono Y, et al.. 2006. Th17
 functions as an osteoclastogenic helper T cell subset that links T cell activation and
 bone destruction. *J Exp Med* 203:2673–2682.

Schillinger U, Lücke FK. 1989. Antibacterial activity of lactobacillus sake isolated from meat.
Appl Environ Microbiol 55:1901-1906.

Seddik HA, Bendali F, Gancel F, Fliss I, Spano G, Drider D. 2017. *Lactobacillus plantarum*
 and its probiotic and food potentialities. *Probiotics Antimicrob Proteins* 9:111-122.

Szulc P, Delmas PD. 2008. Biochemical markers of bone turnover: Potential use in the
 investigation and management of postmenopausal osteoporosis. *Osteoporos Int*
 19:1683-1704.

Turner AS. 2001. Animal models of osteoporosis--necessity and limitations. *Eur Cell Mater*
 1:66-81.

Figure Legend

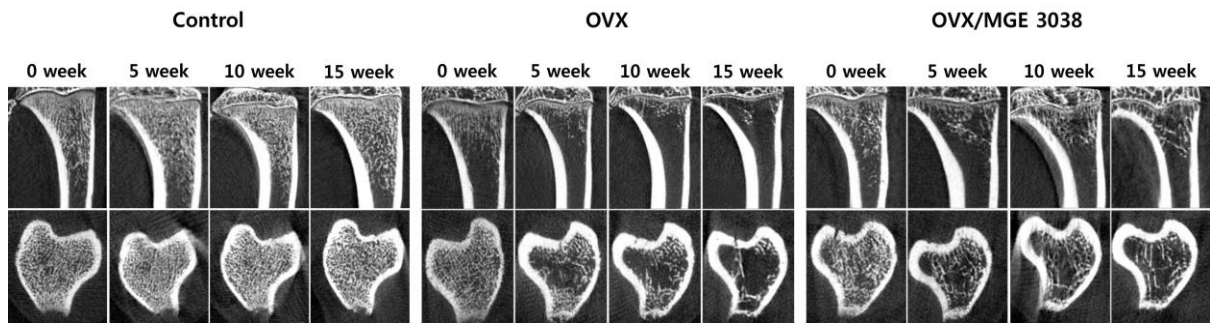


Fig. 1. A micro-computed tomography findings. Although the osteoporotic changes in the ovariectomy-induced osteoporosis group (OVX) and OVX/*Lactiplantibacillus plantarum* MGE 3038 (MGE 3038) feeding group, the preservation of trabecular structure was better in the OVX/MGE 3038 than in OVX group.

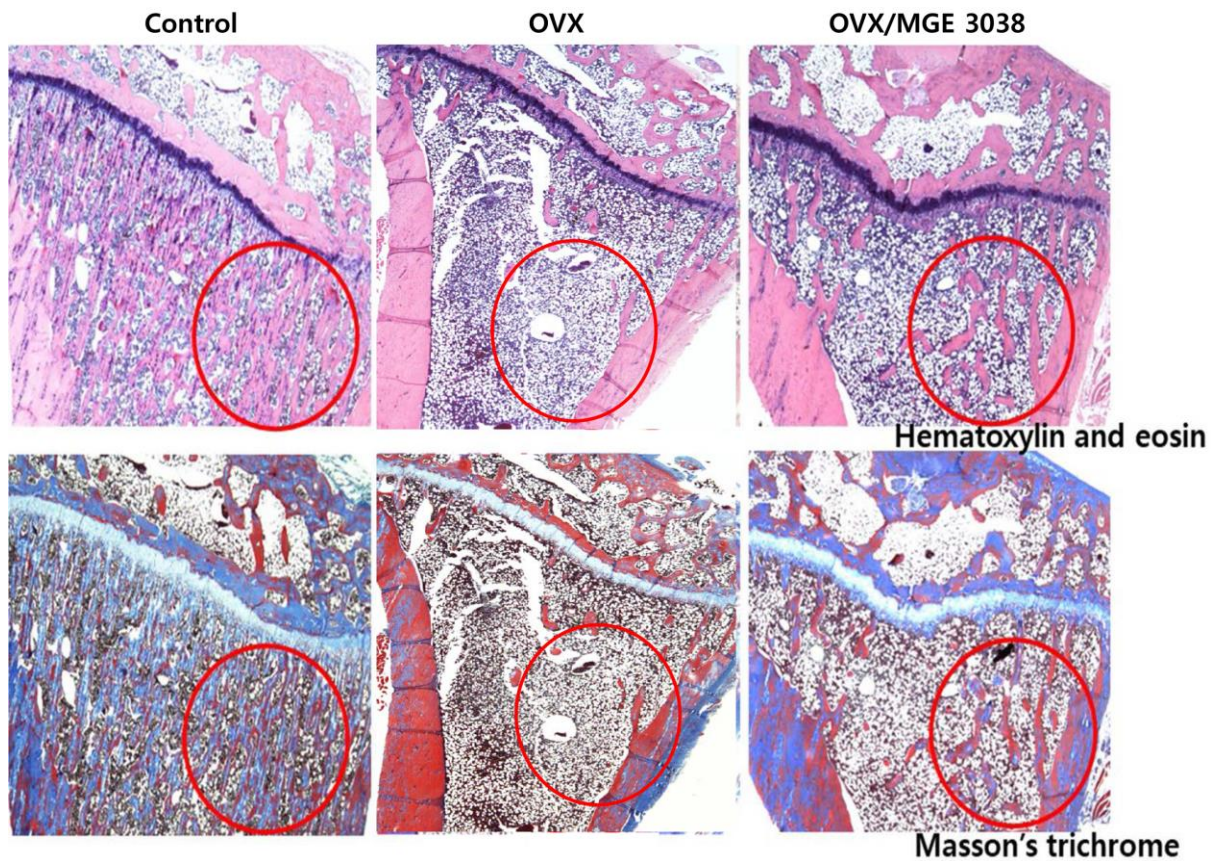
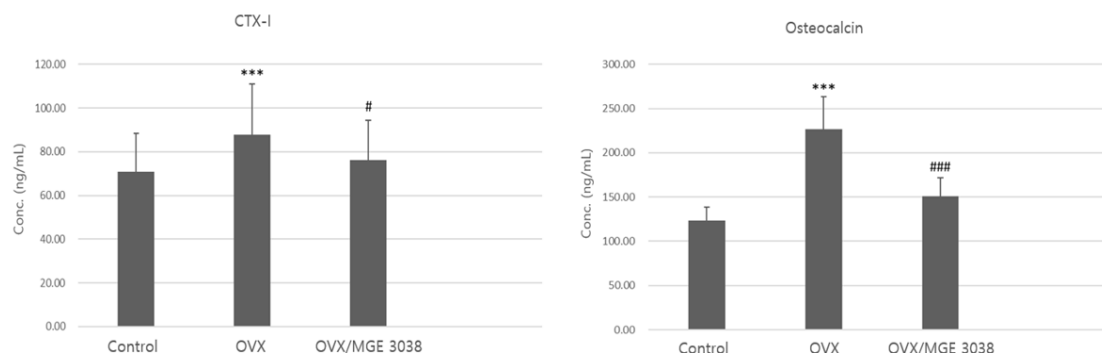


Fig. 2. Pathologic findings of H (hematoxylin) & E (eosin) and Masson's trichrome staining of tibia. The maintenance of trabecular bone structure was seen in the ovariectomy induced osteoporosis (OVX) group and *Lactiplantibacillus plantarum* MGE 3038 feeding group (OVX/MGE 3038) compared to the OVX group.



354

355 Fig. 3. Changes in the levels of serum markers. There were significant differences in serum
 356 levels of C-telopeptide of type I collagen (CTX-I) and osteocalcin between the sham group
 357 (control) and ovariectomy-induced osteoporosis group (OVX) ($p < 0.05$). However, there was
 358 no difference between the control and OVX and *Lactiplantibacillus plantarum* MGE 3038
 359 feeding group (OVX/MGE 3038) ($p > 0.05$). There was also significant difference between the
 360 OVX and OVX/MGE 3038 groups ($p < 0.05$). One way analysis of variance Tukey test. All data
 361 are stated as the mean \pm standard deviation. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$ vs. control. # $p < 0.05$,
 362 ## $p < 0.01$, ### $p < 0.001$ vs. OVX.