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9                   **Effects of Nitrite and Phosphate Replacements for Clean-Label**  
10   **Ground Pork Products**

11  
12   **Abstract**

13   We investigated the effects of different phosphate replacements on the quality of ground pork  
14   products cured with sodium nitrite or radish powder to determine their potential for achieving  
15   clean-label pork products. The experimental design was a  $2 \times 5$  factorial design. For this  
16   purpose, the ground meat mixture was assigned into two groups, depending on nitrite source.  
17   Each group was mixed with 0.01% sodium nitrite or 0.4% radish powder together with 0.04%  
18   starter culture, and then processed depending on phosphate replacement (with or without 0.5%  
19   sodium tripolyphosphate; STPP (+), STPP (-), 0.5% oyster shell calcium (OSC), 0.5% citrus  
20   fiber (CF), or 0.5% dried plum powder (DPP)). All samples were cooked, cooled, and stored  
21   until analysis within two days. The nitrite source had no effect on all dependent variables of  
22   ground pork products. However, in phosphate replacement treatments, the STPP (+) and OSC  
23   treatments had a higher cooking yield than the STPP (-), CF, or DPP treatments. OSC treatment  
24   was more effective for lowering total fluid separation compared to STPP (-), CF, or DPP  
25   treatments, but had a higher percentage than STPP (+). The STPP (+) treatment did not differ  
26   from the OSC or CF treatments for CIE L\* and CIE a\* values. Moreover, no differences were  
27   observed in nitrosyl hemochrome content, lipid oxidation, hardness, gumminess, and  
28   chewiness between the OSC and STPP (+) treatments. In conclusion, among the phosphate  
29   replacements, OSC addition was the most suitable to provide clean-label pork products cured  
30   with radish powder as a synthetic nitrite replacer.

31   **Keywords:** nitrite replacement, phosphate replacement, radish powder, pork products, clean-  
32   label

### 33 **Introduction**

34 As a curing ingredient in meat products, nitrite plays a key role in curing meat color, while  
35 conferring antimicrobial and antioxidant protection, and a curing flavor (Alahakoon et al.,  
36 2015, Pegg and Shahidi, 2000). Despite the benefits of nitrite in meat curing, increasing  
37 consumer awareness of health-related risks associated with synthetic food additives (Hur et al.,  
38 2015) has boosted the demand for 'clean-label products,' such as organic, eco-friendly, and  
39 synthetic additive-free products (Maruyama et al., 2021; Yong et al., 2021). In response to this  
40 need, the meat industry uses pre-conversion of nitrite from vegetable powders or nitrate-rich  
41 vegetable sources together with starter culture is applied to meat products (Jeong, 2016).

42 Celery juice powder, which is widely used, is a feasible alternative to synthetic nitrite.  
43 However, excessive addition of celery juice or powder affects the sensory properties of the  
44 products negatively (Alahakoon et al., 2015), and may cause allergic reactions (Ballmer-Weber  
45 et al., 2002). Therefore, other natural sources, such as vegetables, fruits, and their by-products,  
46 have been studied as alternative nitrite sources. Thus, Riel et al. (2017) found that the addition  
47 of parsley-extract powder to mortadella sausages produced a redness similar to that obtained  
48 by addition of synthetic nitrite. Similarly, Šojića et al. (2020) reported that a mixture of tomato  
49 peel extract and peppermint oil could be used for partial replacement of sodium nitrite in pork  
50 sausages. Moreover, testing different vegetable (Chinese cabbage, radish, and spinach)  
51 powders for nitrite substitution, Jeong et al. (2020) found that the use of radish powder  
52 conferred similar qualities to those obtained by the addition of synthetic nitrite, suggesting its  
53 potential as a substitute for synthetic nitrite. However, for 'clean-label' meat products, other  
54 challenges are faced, and solutions to replace synthetic phosphates are emerging in the meat  
55 industry (Thangavelu et al., 2019).

56 Phosphate is widely used for meat production because of its many functions, including

57 increasing water-holding capacity, inhibiting lipid oxidation, and improving textural and  
58 sensory attributes (Long et al., 2011; Thangavelu et al., 2019). Recognized as a GRAS  
59 (Generally Recognized as Safe) substance by the FDA (Food and Drug Administration),  
60 phosphate can be added at a concentration of 0.5% or less of the final meat products (USDA-  
61 FSIS, 2015). With respect to replacing synthetic phosphates, the use of calcium powders from  
62 natural sources (Bae et al., 2017; Cho et al. 2017), polysaccharides (Meyer, 2018; Öztürk-  
63 Kerimoğlu and Serdaroğlu, 2019), amino acids (Kim et al., 2014), protein hydrolyzates  
64 (Shahidi and Synowieki, 1997; Vann and DeWitt; 2007), dietary fiber (Powell et al., 2019;  
65 Magalhães et al., 2020), and mushrooms (Choe et al., 2018), has been tested. Thus, Bae et al.  
66 (2017) reported that pork meat products containing oyster shell calcium had a texture similar  
67 to that of obtained upon sodium tripolyphosphate treatment. Fernández-Ginés et al. (2003)  
68 reported that Bologna sausages treated with citrus fiber had a cooking yield and emulsion  
69 stability similar to those of products added with sodium tripolyphosphate. Similarly, Jarvis et  
70 al. (2012) confirmed that chicken breast fillets marinated by combining plum powder and plum  
71 fiber showed similar quality characteristics to those obtained upon marinating with sodium  
72 tripolyphosphate. Although several studies have reported effective replacement of synthetic  
73 nitrite and phosphate, studies on the replacement of synthetic phosphate in naturally cured meat  
74 products with a vegetable powder have not been reported.

75 Therefore, in this study we compared oyster shell calcium, citrus fiber, and dried plum  
76 powder as candidate natural phosphate sources for phosphate replacement in meat products  
77 cured with either sodium nitrite or with radish powder as a natural nitrite alternative, aiming to  
78 contribute to the development of clean-label meat production.

79

## 80 **Materials and Methods**

### 81 **Preparation of radish powder and other materials**

82 Fresh radishes (*Raphanus sativus* L.) grown in South Korea were purchased and randomly  
83 selected to manufacture radish powder. Radish powder was prepared after subsequent washing,  
84 homogenizing, drying, and powdering as previously described by Bae et al. (2020). Then,  
85 powdered samples were vacuum-packed and stored at  $-18^{\circ}\text{C}$  until further use. To standardize  
86 the nitrate content (32,000 ppm) from each batch, the radish powder was mixed with  
87 maltodextrin (#186785579, ESfood, Gunpo, Korea) before processing the meat products.

88 A starter culture (Bactoform<sup>®</sup> CS-300, CHR Hansen, Pohlheim, Germany) comprising  
89 *Staphylococcus carnosus* and *Staphylococcus carnosus* subsp., sodium nitrite (S225, Sigma-  
90 Aldrich, St. Louis, MO, USA), sodium tripolyphosphate (238503, Sigma-Aldrich, St. Louis,  
91 MO, USA), sodium chloride (S-3160-65, Fisher Scientific UK, Loughborough, UK), sodium  
92 ascorbate (#35268, Acros Organics, Geel, Belgium), and dextrose (A16828, Thermo Fisher  
93 Scientific, Heysham, UK) were purchased from commercial suppliers. As alternatives to  
94 synthetic phosphate, oyster shell calcium (Glucan Corp., Jinju, Korea), citrus fiber (CF-100,  
95 Fiberstar, Inc., River Falls, WI, USA), and dried plum powder (#80276308572, Sunsweet  
96 Growers Inc., Yuba City, CA, USA) were obtained.

### 97 **Preparation of ground pork products**

98 Fresh pork ham and back fat were purchased from a local market. After trimming  
99 intermuscular fat and visible connective tissues, the lean pork meat and back fat were stored at  
100  $-18^{\circ}\text{C}$  until processing within one month. Frozen materials (total batch size of 35 kg per trial)  
101 were completely thawed and then ground using a chopper (TC-22 Elegant plus, Tre Spade,  
102 Torino, Italy) equipped with a 3-mm plate. Ground mixtures were randomly divided into ten  
103 portions and assigned to two groups (five batches each) depending on the nitrite source (Table

104 1). First, 70% pork meat and 15% back fat were mixed for 3 min with 0.01% sodium nitrite or  
105 0.4% radish powder and 0.04% starter culture in a mixer (5K5SS, Whirlpool, St. Joseph, MI,  
106 USA). Second, each group was processed depending on phosphate replacement, including with  
107 or without 0.5% sodium tripolyphosphate (STPP) or 0.5% phosphate replacement (oyster shell  
108 calcium; OSC, citrus fiber; CF, and dried plum powder; DPP). Other ingredients (1.5% sodium  
109 chloride, 1% dextrose, and 0.05% sodium ascorbate; total meat mixture basis) along with 15%  
110 ice/water were added to a mixer and mixed again for 7 min. The treatments were filled into 50  
111 mL conical tubes. Five batches of sodium nitrite were placed in a refrigerator at 4°C for 1 h.  
112 The remaining five batches of radish powder and starter culture were stored in an incubator at  
113 40°C for 2 h to allow the conversion of nitrate to nitrite. All samples were cooked to 75°C in a  
114 water bath (MaXturdy 45, Daihan Scientific, Wonju, Korea) at 90°C. Once cooking, the  
115 samples were cooled for 20 min in ice slurry and stored overnight at 2–3°C in the dark until  
116 analysis. Experiments were performed in triplicate, and all dependent variables were measured  
117 in duplicate.

#### 118 **Determination of pH and cooking yield**

119 The pH was measured with a pH meter (Accumet AB150, Thermo Fisher Scientific, Inc.,  
120 Singapore) after adding 25 mL of distilled water to a 5 g sample and homogenized (DI-25 basic,  
121 IKA®-Werke Gmb & Co. KG, Staufen, Germany). Five samples per each batch were weighed  
122 before cooking and after cooking and cooling overnight. Cooking yield was calculated as  
123 follows: [cooked sample weight/raw sample weight] × 100.

#### 124 **Total fluid, lipid, and water separation**

125 Total fluid, lipid, and water separation of ground pork products was measured by the method  
126 described by Hughes et al. (1997) and Lee et al. (2008). Twenty grams of the uncooked meat

127 mixture was placed into a 50-mL conical tube with a mesh. After weight measurement, the  
128 conical tubes filled with the samples were cooked for 30 min in a water bath at 75°C (CB60L,  
129 Dongwon Scientific Instrument, Busan, Korea), cooled for 20 min, and centrifuged at 500 × g  
130 for 5 min. Pellets and supernatants in the conical tubes were weighed before drying. The  
131 supernatant was dried for 18 h at 105°C using a dryer (ON-12GW; Jeio Tech Co. Ltd., Daejeon,  
132 Korea) and weighed again. The percentage total fluid separation (TFS), lipid separation (LS),  
133 and water separation (WS) were calculated using the following equations:

$$\% \text{ TFS} = \frac{\text{Weight of sample before cooking (g)} - \text{Weight of pellet after cooking and centrifuging (g)}}{\text{Weight of sample before cooking (g)}} \times 100$$

$$\% \text{ LS} = \frac{\text{Weight of dried supernatant (g)}}{\text{Weight of sample before cooking (g)}} \times 100$$

$$\% \text{ WS} = \% \text{ TFS} - \% \text{ LS}$$

### 137 **Color measurements**

138 After cutting the samples in the longitudinal direction, the cut surfaces of samples were  
139 measured for CIE (the International Commission on Illumination ) L\*a\*b\* values using a  
140 colorimeter (CR-400, 8 mm aperture, illuminant C, 2° standard observer; Konica Minolta  
141 Sensing Inc., Osaka, Japan) after calibrating the standard plate (L\* 94.87, a\* -0.39, b\* 3.88).  
142 Two readings were recorded on each cut surface for each pork sausage immediately after  
143 cutting.

### 144 **Nitrosyl hemochrome and 2-thiobarbituric acid-reactive substances (TBARS)**



145 **determination**

146 Nitrosyl hemochrome in pork products was measured using the method described by  
147 Hornsey (1956). After extraction and filtration, absorbance of the filtrate was determined at  
148 540 nm ( $A_{540}$ ) using a spectrophotometer (UV-1800, Shimadzu Corp., Kyoto, Japan). Nitrosyl  
149 hemochrome concentration (ppm) was calculated by multiplying absorbance ( $A_{540}$ ) by 290.  
150 TBARS values was measured using the method described by Tarladgis et al. (1960). Briefly,  
151 after reacting malondialdehyde (MDA) in samples with 0.02 M 2-thiobarbituric acid (TBA)  
152 solution, absorbance of reactive substances was determined at 538 nm. The results were  
153 multiplied by a factor of 7.8 to calculate TBARS values (mg MDA/kg samples).

154 **Texture profile analysis**

155 After cutting the cross section of the samples to a thickness of 2.5 cm, the hardness,  
156 springiness, cohesiveness, gumminess, and chewiness of the samples (2.8 cm diameter) were  
157 measured using a texture analyzer (TA-XT2i, Stable Micro Systems, Surrey, UK) equipped  
158 with a 50-mm aluminum cylinder. Crosshead speed for the measurements was 5 mm/s and  
159 compression was 40% of sample thickness.

160 **Statistical analysis**

161 The experimental design was a  $2 \times 5$  factorial design with two nitrite sources (sodium nitrite  
162 or radish powder) and five phosphate replacement treatments (with or without phosphate,  
163 oyster shell calcium, citrus fiber, or dried plum powder). All data were statistically analyzed  
164 using the PROC GLIMMIX procedure in the SAS software (version 9.4; SAS, 2012) to  
165 determine fixed effects for nitrite and phosphate replacement and their interactions. When  
166 significance ( $p < 0.05$ ) was determined, the least squares means were further separated using the  
167 LINES option in the same software.

168

## 169 **Results and Discussion**

170 The significance of nitrite sources (N), phosphate replacements (P), and their interaction was  
171 shown in Table 2. A two-way interaction ( $N \times P$ ) between the main effects was not found  
172 ( $p > 0.05$ ) for any dependent variables tested in this study. Therefore, the results for individual  
173 main effects are presented.

### 174 **pH**

175 Nitrite sources (N) did not affect ( $p > 0.05$ ) the pH of pork products (Table 2), indicating that  
176 there were no significant ( $p > 0.05$ ) differences in pH between sodium nitrite- and radish  
177 powder-treated pork products (Table 3). These findings agreed with those reported by Sindelar  
178 et al. (2007) and Yoon et al. (2021), who found that pH of meat products naturally cured with  
179 celery juice powder and white kimchi powder, respectively, did not differ from those of meat  
180 products cured with sodium nitrite. In contrast, phosphate replacements (P) was found to  
181 significantly ( $p < 0.001$ ) affect the pH of ground pork products (Tables 2). Thus, the OSC  
182 treatment had the highest ( $p < 0.05$ ) pH values, while the CF and DPP treatments had lower  
183 ( $p < 0.05$ ) pH values than either the STPP (+) or STPP (-) treatments (Table 3). In our  
184 preliminary test, the pH of OSC was 9.93, whereas those of CF and DPP ranged from 3.60 to  
185 4.05. It is likely that organic acids, such as citric acid, quinic acid, and malic acid contained  
186 in citrus fiber and dried plum powder reduced the pH of the final products (Bae et al., 2014;  
187 Song et al., 1998).

### 188 **Cooking yield**

189 Cooking yield was not affected ( $p > 0.05$ ) by nitrite sources (N) (Tables 2 and 3). Conversely,  
190 Yoon et al. (2021) showed that pork sausages containing sodium nitrite showed a higher

191 cooking yield than those containing white kimchi powder. However, Jeong et al. (2020) found  
192 no significant difference in cooking yield between pork products cured with various vegetable  
193 powders (Chinese cabbage, radish, and spinach) and sodium nitrite-added products,  
194 consistently with the findings reported herein. However, in this study, phosphate replacements  
195 (P) had a significant ( $p < 0.001$ ) effect on cooking yield of ground pork products (Table 2).  
196 Interestingly, the cooking yield in the OSC treatment was 96.81%, which did not differ  
197 significantly ( $p > 0.05$ ) from that of the STPP (+) treatment (98.45%) (Table 3). Similarly, Lee  
198 et al. (2011) found that emulsion-type pork sausages treated with 0.3% STPP showed similar  
199 cooking yield as those treated with 0.5% oyster shell powder. However, the CF and DPP  
200 treatments showed a lower ( $p < 0.05$ ) cooking yield than the STPP (+) treatment (Table 3).  
201 Dietary fiber from citrus fruits and sorbitol in dried plum powder have been introduced as  
202 good candidates for improving the water retention capacity of meat systems (Fernández-Ginés  
203 et al., 2003; Jarvis et al., 2015; Lundberg et al., 2014). However, the unexpected results for  
204 the CF and DPP treatments in this study might be attributed to the fact that the organic acids  
205 contained in CF and DPP had a negative effect on the cooking yield of ground pork products.  
206 Consistently, with regard to the effect of organic acids on meat products, Bae et al. (2021)  
207 reported that naturally cured sausages containing more organic acids showed a lower pH,  
208 thereby resulting in a lower cooking yield, which supports our findings.

### 209 **Total fluid separation (TFS), lipid separation (LS), and water separation (WS)**

210 Neither TFS, LS, nor WS of pork products were affected ( $p > 0.05$ ) by nitrite sources (N)  
211 (Tables 2 and 3). However, significant ( $p < 0.001$ ) phosphate replacement (P) effects were  
212 observed for TFS, LS, and WS in ground pork products (Table 2). The OSC treatment had a  
213 significantly ( $p < 0.05$ ) higher TFS than the STPP (+) treatment, but lower ( $p < 0.05$ ) than the  
214 STPP (-), CF, and DPP treatments (Table 3). Among the phosphate replacement treatments,

215 the low TFS of the OSC treatment may be due to the increased water holding capacity owing  
216 to the high pH of OSC itself (Park, 2011). The highest TFS ( $p < 0.05$ ) was seen in the CF  
217 treatment. Since dietary fiber has a microporous structure, it can adsorb moisture and fat, but  
218 it is thought that the adsorbed components were discharged by strong physical forces such as  
219 centrifugation during the experiment (Lundberg et al., 2014; Wang et al., 2015). A lower TFS  
220 was observed with DPP treatment than with CF treatment in this study, likely because the high  
221 sorbitol content of DPP may affect its moisture binding ability (Javis et al., 2015). However,  
222 there were no significant differences ( $p > 0.05$ ) in LS between the OSC and STPP (+)  
223 treatments (Table 3). In addition, these treatments had a significantly ( $p < 0.05$ ) lower LS than  
224 the CF or the DPP treatments. Similar to the TFS results described above, the same trend was  
225 observed for WS (Table 3). The greatest WS ( $p < 0.05$ ) was observed in the CF treatment, likely  
226 due to the coalescence and agglomeration of fiber particles (Powell et al., 2019). Overall, our  
227 results suggest that, among the phosphate replacements tested in this study, OSC has the  
228 potential to substitute STPP in ground pork products in terms of water-holding capacity,  
229 regardless of the nitrite source.

### 230 **CIE color**

231 The nitrite sources (N) had no effects ( $p > 0.05$ ) on the CIE L\* values of cooked products  
232 (Tables 2 and 4). Similarly, Choi et al. (2020) found that pork sausages cured with white  
233 kimchi powder obtained similar CIE L\* values as those cured with sodium nitrite, although  
234 the type of vegetable powder used was different from that in this study. However, phosphate  
235 replacements (P) did significantly ( $p < 0.001$ ) affect CIE L\* values of cooked products (Table  
236 2). The OSC treatment did not differ ( $p > 0.05$ ) in CIE L\* values from STPP (+) and CF  
237 treatments, but lower ( $p < 0.05$ ) than those in the STPP (-) treatment (Table 4). DPP treatment  
238 obtained the lowest CIE L\* values ( $p < 0.05$ ). Similarly, Lee and Ahn (2005) observed that the

239 inclusion of plum extract in turkey breast rolls resulted in reduced L\* values.

240 The CIE a\* values of products containing sodium nitrite and radish powder were 9.87 and  
241 9.86, respectively, and were not significantly ( $p>0.05$ ) affected by nitrite sources (N) (Tables  
242 2 and 4). Similar results were obtained by Bae et al. (2020), who reported that pork products  
243 cured with radish powder showed CIE a\* values similar to those with sodium nitrite. Yoon et  
244 al. (2021) also found that there were no significant differences in CIE a\* values between pork  
245 sausages added with sodium nitrite and those added with white kimchi powder. Additionally,  
246 the main effect of phosphate replacement (P) on CIE a\* values was significant ( $p<0.001$ ; Table  
247 2). The CIE a\* values were lowest ( $p<0.05$ ) for the DPP treatment and did not significantly  
248 ( $p>0.05$ ) differ among treatments (Table 4). These CIE a\* values were in agreement with those  
249 of Meyer (2018), who reported that the addition of plum concentrate as a phosphate  
250 replacement in whole muscle hams resulted in a decrease in redness.

251 Nitrite sources (N) did not affect ( $p>0.05$ ) CIE b\* values of ground pork products (Tables 2  
252 and 4). Jeong et al. (2020) found that cooked meat products added with 0.4% radish powder  
253 showed similar CIE b\* values to those with 150 ppm sodium nitrite, as shown in this study.  
254 Overall, our CIE color results suggest that radish powder is a useful alternative to synthetic  
255 nitrite for clean-label meat products. However, phosphate replacements (P) significantly  
256 ( $p<0.001$ ) affected CIE b\* values of ground pork products (Table 2). All phosphate  
257 replacement treatments significantly ( $p<0.05$ ) increased the CIE b\* values, compared to the  
258 STPP (+) treatment (Table 4). The impact of the addition of OSC on CIE b\* of ground pork  
259 products was smaller, although significant. In contrast, DPP treatment showed the highest CIE  
260 b\* values ( $p<0.05$ ), probably due to the color of the endogenous pigments in the plant extract  
261 (Nowak et al., 2016; Riel et al., 2017). Thus, the use of DPP may have a negative effect on  
262 the color of ground pork products.

## 263 **Nitrosyl hemochrome**

264 Nitrosyl hemochrome, which provides a typical cured-meat color, is formed by the reaction  
265 of myoglobin with nitric oxide reduced from nitrite during cooking (Parthasarathy and Bryan,  
266 2012). In this study, nitrite sources (N) had no effect ( $p>0.05$ ) on nitrosyl hemochrome content  
267 in ground pork products (Tables 2 and 4), indicating that radish powder is a good candidate as  
268 a synthetic nitrite substitute for cured meat color. However, nitrosyl hemochrome content was  
269 significantly ( $p<0.001$ ) affected by phosphate replacements (P) (Table 2). CF and DPP  
270 treatments had significantly ( $p<0.05$ ) higher nitrosyl hemochrome contents than STPP (-),  
271 STPP (+), or OSC treatments, and the highest ( $p<0.05$ ) nitrosyl hemochrome content was  
272 observed in DPP treatment (Table 4). As a decrease in pH can promote the rate of the curing  
273 reaction (Honikel, 2008), organic acids and polyphenols in CF or DPP may accelerate meat  
274 curing by lowering the pH or acting as a reducing agent (Ahmad et al., 2015; Terns et al.,  
275 2011). In contrast, the OSC treatment had the lowest ( $p<0.05$ ) nitrosyl hemochrome content  
276 across all treatments, probably due to the high pH of OSC powder used in this study (Table  
277 4). Nevertheless, in this study, there was no difference in CIE  $a^*$  value in OSC treatment and  
278 other treatments except for DPP treatment. This may be because the high pH of OSCs limited  
279 the curing process or the denaturation of myoglobin (Honikel, 2008; Trout, 1989).

## 280 **2-thiobarbituric acid-reactive substances (TBARS)**

281 The TBARS values were not significantly ( $p>0.05$ ) influenced by nitrite sources (N) or  
282 phosphate replacements (P) in ground pork products (Table 2). Regardless of nitrite sources,  
283 TBARS values of all cooked products were 0.12 mg MDA/kg (Table 4). These findings agreed  
284 with those reported by Magrinyà et al. (2016), who found that cooked cured sausages had  
285 similar TBARS values despite different nitrite treatment (sodium nitrite or vegetable powder).  
286 It is likely that nitrites reduced from nitrates contained in radish powder as well as antioxidants

287 present in radish inhibited lipid rancidity (Ahn et al, 2019; Ozaki et al., 2021). Thus, adding  
288 radish powder may have a similar inhibitory effect on lipid oxidation as that which results  
289 from treatment with synthetic nitrite. Furthermore, TBARS values of ground pork products  
290 were not significantly ( $p>0.05$ ) affected by phosphate replacements (P) (Tables 2 and 4). This  
291 result is supported by previous research on substitution for synthetic phosphates. Lee et al.  
292 (2011) reported that OSC had similar efficacy in inhibiting lipid oxidation as STPP in  
293 emulsion-type sausages. Powell et al. (2019) found that Bologna sausages treated with citrus  
294 fibers did not differ in TBARS value from those treated with STPP. Moreover, Nuñez de  
295 Gonzlaes et al. (2009) obtained similar TBARS values when dried plum concentrate or  
296 phosphate was added to boneless ham.

### 297 **Textural properties**

298 Nitrite sources (N) had no significant ( $p>0.05$ ) effect on the textural properties of ground  
299 pork products (Tables 2 and 5). Our results were supported by Sucu and Turp (2018), who  
300 reported no difference in texture profile between nitrite- and beetroot powder-added Turkish  
301 fermented sausages. However, significant effects of phosphate replacements (P) on ground  
302 pork products was observed ( $p<0.001$ ) only for cohesiveness and springiness (Table 2),  
303 whereas neither hardness, gumminess, nor chewiness were affected ( $p>0.05$ ) by phosphate  
304 replacement. The STPP (+) treatment showed the highest cohesiveness and springiness  
305 ( $p<0.05$ ), while cohesiveness and springiness of STPP (-), CF, and DPP treatments were lower  
306 ( $p<0.05$ ) than those of STPP (+) and OSC treatments, but were similar to each other ( $p>0.05$ )  
307 (Table 5). Consistently with the findings reported herein, recently, Lee (2020) reported that  
308 the addition of OSC resulted in higher cohesiveness, springiness, and chewiness than  
309 restructured hams containing STPP, although, in our study, chewiness did not differ. Similarly,  
310 Powell et al. (2019) found that bologna sausages added with 0.5% citrus fiber had lower

311 cohesiveness and springiness than those added with STPP but, in agreement with our results,  
312 hardness, gumminess, and chewiness were similar between them. However, Lee and Ahn  
313 (2005) reported that the hardness, cohesiveness, springiness, and chewiness of turkey breast  
314 rolls were not influenced by the addition of up to 3% plum extract, which partially agrees with  
315 our results. Consequently, the addition of OSC resulted in lower cohesiveness and springiness  
316 of ground pork products, compared to synthetic phosphate, although OSC can have a greater  
317 effect on the texture of the final products among the phosphate replacement treatments tested  
318 here, as observed by Bae et al. (2017) for various calcium powders.

319

## 320 **Conclusion**

321 Nitrite sources (sodium nitrite or radish powder) did not significantly affect the  
322 physicochemical or textural properties of ground pork products. However, most dependent  
323 variables were influenced by phosphate replacement treatment. The addition of oyster shell  
324 calcium maintained cooking yield and lipid separation, replacing sodium tripolyphosphate in  
325 the final products. In contrast, ground pork products with citrus fiber or dried plum powder  
326 showed a negative effect on water and lipid binding ability. In particular, the addition of dried  
327 plum powder resulted in a difference in color in ground pork products compared to STPP (+)  
328 treatment. Pork products with oyster shell calcium showed textural properties relatively  
329 similar to those of products treated with sodium tripolyphosphate. Therefore, oyster shell  
330 calcium is suitable as a synthetic phosphate substitute for clean-label ground pork products  
331 when cured with radish powder.

332

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478

479 **Table 1. Experimental design (2 × 5 factorial) to investigate the effects of nitrite and**  
 480 **phosphate replacements for ground pork products**

Samples	Nitrite sources <sup>1</sup>	Phosphate replacements <sup>2</sup>
1	Sodium nitrite	No sodium tripolyphosphate
2	Sodium nitrite	Sodium tripolyphosphate
3	Sodium nitrite	Oyster shell calcium
4	Sodium nitrite	Citrus fiber
5	Sodium nitrite	Dried plum powder
6	Radish powder	No sodium tripolyphosphate
7	Radish powder	Sodium tripolyphosphate
8	Radish powder	Oyster shell calcium
9	Radish powder	Citrus fiber
10	Radish powder	Dried plum powder

481 <sup>1</sup> Nitrite sources: Two different nitrite sources (NaNO<sub>2</sub> or radish powder) were used. Radish powder  
 482 was added with a starter culture comprising *S. carnosus* and *S. carnosus* subsp.

483 <sup>2</sup> Phosphate replacement treatments: Samples prepared with or without 0.5% sodium tripolyphosphate  
 484 or with one of three different synthetic phosphate replacements (0.5% oyster shell calcium: OSC, citrus  
 485 fiber: CF, or dried plum powder; DPP).

486 **Table 2. Significance of main and interaction effects on nitrite sources and phosphate replacements on physicochemical properties of ground**  
 487 **pork products**

Main and interaction effects <sup>1</sup>	Dependent variables <sup>2</sup>														
	pH	Cooking yield	TFS	LS	WS	CIE L*	CIE a*	CIE b*	Nitrosyl hemochrome	TBARS	Hardness	Cohesiveness	Springiness	Gumminess	Chewiness
Nitrite sources <sup>3</sup> (N)	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS
Phosphate replacements <sup>4</sup> (P)	**	**	**	**	**	**	**	**	**	NS	NS	**	**	NS	NS
N × P	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS

488 <sup>1</sup> Main and interaction effects: \* = p<0.05, \*\* = p<0.001, NS = not significant.

489 <sup>2</sup> Dependent variables: TFS (total fluid separation), LP (lipid separation), WP (water separation), CIE L\* (lightness), CIE a\* (redness), CIE b\* (yellowness), and TBARS  
 490 (2-thiobarbituric acid reactive substances).

491 <sup>3</sup> Nitrite sources: Two different nitrite sources (NaNO<sub>2</sub> or radish powder) were used. Radish powder was added with a starter culture comprising *S. carnosus* and *S.*  
 492 *carnosus* subsp.

493 <sup>4</sup> Phosphate replacement treatments: Samples prepared with or without 0.5% sodium tripolyphosphate or with one of three different synthetic phosphate replacements  
 494 (0.5% oyster shell calcium; OSC, citrus fiber; CF, or dried plum powder; DPP).

495  
 496



497 **Table 3. Effects of nitrite and phosphate replacements on pH, cooking yield, total fluid separation, lipid separation, and water separation in**  
 498 **ground pork products**

Main effects	pH	Cooking yield (%)	Total fluid separation (%)	Lipid separation (%)	Water separation (%)
Nitrite sources <sup>1</sup> (N)					
Sodium nitrite	6.25	93.39	11.14	1.01	10.13
Radish powder	6.25	93.02	11.24	1.03	10.21
SEM	0.01	1.27	0.47	0.11	0.36
Phosphate replacements <sup>2</sup> (P)					
STPP (-)	6.09 <sup>C</sup>	91.06 <sup>B</sup>	13.03 <sup>C</sup>	1.20 <sup>B</sup>	11.83 <sup>C</sup>
STPP (+)	6.32 <sup>B</sup>	98.45 <sup>A</sup>	5.08 <sup>E</sup>	0.25 <sup>C</sup>	4.83 <sup>E</sup>
OSC	6.79 <sup>A</sup>	96.81 <sup>A</sup>	6.83 <sup>D</sup>	0.42 <sup>C</sup>	6.41 <sup>D</sup>
CF	6.03 <sup>D</sup>	89.76 <sup>B</sup>	16.62 <sup>A</sup>	1.77 <sup>A</sup>	14.85 <sup>A</sup>
DPP	6.04 <sup>D</sup>	89.97 <sup>B</sup>	14.39 <sup>B</sup>	1.46 <sup>AB</sup>	12.93 <sup>B</sup>
SEM	0.01	1.38	0.56	0.14	0.43

499 <sup>1</sup> Nitrite sources: Two different nitrite sources (NaNO<sub>2</sub> or radish powder) were used. Radish powder was added with a starter culture comprising *S. carnosus* and *S.*  
 500 *carnosus* subsp.

501 <sup>2</sup> Phosphate replacement treatments: Samples prepared with or without 0.5% sodium tripolyphosphate or with one of three different synthetic phosphate  
 502 replacements (0.5% oyster shell calcium; OSC, citrus fiber; CF, or dried plum powder; DPP).

503 <sup>A-E</sup> Means within columns followed by different superscript letters are significantly different (p<0.05).

**Table 4. Effects of nitrite and phosphate replacements on CIE color, nitrosyl hemochrome, and TBARS values in ground pork products**

Main effects	CIE L*	CIE a*	CIE b*	Nitrosyl hemochrome (ppm)	TBARS (mg MDA/kg)
Nitrite sources <sup>1</sup> (N)					
Sodium nitrite	67.62	9.87	8.70	36.09	0.12
Radish powder	67.32	9.86	8.66	35.72	0.12
SEM	0.14	0.05	0.04	0.34	0.03
Phosphate replacements <sup>2</sup> (P)					
STPP (-)	68.67 <sup>A</sup>	10.15 <sup>A</sup>	7.67 <sup>C</sup>	36.71 <sup>C</sup>	0.14
STPP (+)	68.18 <sup>AB</sup>	9.93 <sup>A</sup>	6.25 <sup>E</sup>	33.66 <sup>D</sup>	0.13
OSC	68.02 <sup>B</sup>	9.93 <sup>A</sup>	6.75 <sup>D</sup>	32.91 <sup>D</sup>	0.13
CF	68.50 <sup>AB</sup>	10.03 <sup>A</sup>	8.50 <sup>B</sup>	37.82 <sup>B</sup>	0.10
DPP	63.97 <sup>C</sup>	9.27 <sup>B</sup>	14.23 <sup>A</sup>	39.71 <sup>A</sup>	0.09
SEM	0.21	0.08	0.04	0.42	0.03

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<sup>1</sup> Nitrite sources: Two different nitrite sources (NaNO<sub>2</sub> or radish powder) were used. Radish powder was added with a starter culture comprising *S. carnosus* and *S. carnosus* subsp.

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507

<sup>2</sup> Phosphate replacement treatments: Samples prepared with or without 0.5% sodium tripolyphosphate or with one of three different synthetic phosphate replacements (0.5% oyster shell calcium; OSC, citrus fiber; CF, or dried plum powder; DPP).

508

509

<sup>A-E</sup> Means within columns followed by different superscript letters are significantly different (p<0.05).

510

TBARS: 2-thiobarbituric acid reactive substances. MDA: malondialdehyde.

511

512 **Table 5. Effects of nitrite and phosphate replacements on textural properties in ground pork products**

Main effects	Hardness (N)	Cohesiveness	Springiness	Gumminess (N)	Chewiness (N)
Nitrite sources <sup>1</sup> (N)					
Sodium nitrite	34.43	0.74	0.93	25.56	23.75
Radish powder	35.52	0.74	0.92	26.40	24.29
SEM	2.20	0.01	0.01	2.05	2.00
Phosphate replacements <sup>2</sup> (P)					
STPP (-)	34.95	0.72 <sup>C</sup>	0.91 <sup>C</sup>	25.23	22.99
STPP (+)	34.51	0.79 <sup>A</sup>	0.96 <sup>A</sup>	27.28	26.06
OSC	34.10	0.76 <sup>B</sup>	0.93 <sup>B</sup>	25.88	24.13
CF	36.67	0.71 <sup>C</sup>	0.90 <sup>C</sup>	26.78	23.81
DPP	34.65	0.72 <sup>C</sup>	0.91 <sup>C</sup>	25.23	23.12
SEM	2.29	0.01	0.01	2.12	2.07

513 <sup>1</sup>Nitrite sources: Two different nitrite sources (NaNO<sub>2</sub> or radish powder) were used. Radish powder was added with a starter culture comprising *S. carnosus* and *S.*  
514 *carnosus* subsp.

515 <sup>2</sup> Phosphate replacement treatments: Samples prepared with or without 0.5% sodium tripolyphosphate or with one of three different synthetic phosphate  
516 replacements (0.5% oyster shell calcium; OSC, citrus fiber; CF, or dried plum powder; DPP).

517 <sup>A-C</sup> Means within columns followed by different superscript letters are significantly different (p<0.05).