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<b>Running Title (within 10 words)</b>	Pretreatment method for analysis of vitamin B12
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12 **Abstract** Vitamin B<sub>12</sub> deficiency may lead to serious health issues in both infants and  
13 adults. A simple analytical method involving sample pretreatment with enzyme, followed by  
14 cyanide addition under acidic conditions; separation on an immunoaffinity column; and high-  
15 performance liquid chromatography was developed for the rapid detection and quantitation of  
16 vitamin B<sub>12</sub> in powdered milk. Detection limit and powdered milk recovery were determined  
17 by quantitative analysis. The limits of detection and quantitation were 2.71 and 8.21 µg/L,  
18 respectively. Relative standard deviations of the intra-day and inter-day precisions varied in the  
19 ranges of 0.98–5.31% and 2.16–3.90%, respectively. Recovery of the analysis varied in the  
20 range of 83.41–106.57%, suggesting that the values were acceptable. Additionally, vitamin B<sub>12</sub>  
21 content and recovery in SRM 1849a were 54.10 µg/kg and 112.24%, respectively. Our results  
22 suggested that the analytical method, including the sample pretreatment step, was valid. This  
23 analytical method can be implemented in many laboratory-scale experiments that seek to save  
24 time and labor. Therefore, this study shows that immunoaffinity–high-performance liquid  
25 chromatography/ultraviolet is an acceptable technique for constructing a reliable database on  
26 vitamin B<sub>12</sub> in powdered milk containing starch as well as protein and/or fat in high amounts.

27 **Keywords** vitamin B<sub>12</sub>, powdered milk, HPLC, analytical method

28

## 29 **Introduction**

30 Vitamin B<sub>12</sub> is a water-soluble vitamin belonging to a family of compounds called  
31 cobalamins. Amongst the cobalamins, cyanocobalamin, hydroxycobalamin,  
32 adenosylcobalamin, and methylcobalamin are the major forms of vitamin B<sub>12</sub> (Pakin et al.,  
33 2005; Anatol et al., 2019; Cho et al., 2019). Vitamins are produced by microorganisms and are  
34 accumulated in the liver. Thus, they are found in animal products such as meat, fish, egg, and  
35 milk products but are present in vegetables in very low concentrations (ng/g). The  
36 recommended daily intake of vitamin B<sub>12</sub> is 2.4 µg/day for a Korean adult and 2.6 µg/day for  
37 pregnant and lactating women (Choi et al., 2008; Jang et al., 2014; Moon et al., 2018). Although  
38 the recommended value is very low, vitamin B<sub>12</sub> deficiencies have been shown to affect  
39 neurodevelopment in infants. Additionally, vitamin B<sub>12</sub> deficiency may lead to megaloblastic  
40 anemia, nervous system disorders, and/or improper synthesis of DNA (Cho et al., 2019).

41 The recent advances in this field have drawn the consumers' attention to minor nutrients, such  
42 as vitamins. However, there are only limited reliable databases for vitamin B<sub>12</sub> for the  
43 evaluation of national nutrition in South Korea. The complex structure and multiple possible  
44 vitamers render the analysis of vitamin B<sub>12</sub> particularly challenging (Fang et al., 2017).

45 Vitamin B<sub>12</sub> has been analyzed using several methods, including spectrophotometry,  
46 microbiological methods, and high-performance liquid chromatography (HPLC) (Esteve et al.,  
47 2002; Guggisberg et al., 2012). Microbiological assays and chromatographic approaches are  
48 the most suitable methods for determining the vitamin B<sub>12</sub> content in food (Szterk et al., 2012).

49 Microbiological assays are the oldest assay method and the most commonly used technique for  
50 vitamin B<sub>12</sub> detection. Although such assays are highly sensitive, they lack specificity as  
51 inactive cobalamins in some food matrices may interfere with the microorganism growth.  
52 These methods are also time-consuming, as they involve steps such as tissue culture and

53 preservation of strain. Moreover, these methods lack sensitivity and have low precision  
54 (O’Broin and Kelleher, 1992).

55 Numerous methods for the analysis of vitamin B<sub>12</sub> have been described by Karmi et al. (2011).  
56 Among them, HPLC–mass spectrometry is probably the most frequently used technique for  
57 determining vitamin B<sub>12</sub> in food and biological samples. To overcome the low sensitivity of the  
58 existing techniques, which is a limitation, an attempt was made to obtain food samples with  
59 low concentrations of vitamin and analyze them through pretreatment methods such as sample  
60 concentration using solid phase extraction or immunoaffinity columns (Iwase and Ono, 1997;  
61 Heudi et al., 2006; Sun et al., 2016; Jie et al., 2019). Vitamin B<sub>12</sub> exists in free and bound forms  
62 in foods. It can be extracted from protein-rich foods using proteolytic enzymes. However,  
63 information on the extraction of vitamin B<sub>12</sub> from powdered milk is very limited. Especially,  
64 powdered milk add starch as well as protein and/or fat to improve its nutrition value (Seo et al.,  
65 2018). The presence of these additional components renders the analysis of vitamin B<sub>12</sub>  
66 extremely difficult (Lee et al., 2015; Bito et al., 2016). Hence, the analysis of vitamin B<sub>12</sub> in  
67 powder milk must include a pretreatment step. Currently, the methods validated by the Ministry  
68 of Food and Drug Safety (MFDS) apply to infant formula, baby formula diet, and milk formulas;  
69 however, powdered milk containing starch is not included in this list. In this study, a  
70 chromatographic approach involving a pretreatment step and immunoaffinity column  
71 purification during the sample preparation of powdered milk containing starch was adopted to  
72 remove interfering matrix components and enrich the sample with the target analyte to ease  
73 quantitation. This analytical method involving a pretreatment step coupled with  
74 immunoaffinity purification and HPLC/Ultraviolet (UV) was validated and applied for the  
75 determination of total vitamin B<sub>12</sub> content in powdered milk containing starch.

76

## 77 **Materials and Methods**

### 78 **Standard, sample, and reagent**

79 The powdered milk used in this study was purchased from a local market and kept at 4°C  
80 for further use. An powdered milk standard reference material, SRM 1849a (National Institute  
81 of Standard and Technology, USA), which is a certified reference material, was used in the  
82 recovery tests. Vitamin B<sub>12</sub> content in SRM 1849a was 48.2±8.5 µg/kg. Sodium acetate was  
83 purchased from Junsei Chemical (Japan), while the enzyme, amylase, was purchased from  
84 ANKOM (catalogue TAHTL-NC24). HPLC grade water and acetonitrile were purchased from  
85 Merck (Germany).

### 87 **Preparation of standards**

88 Vitamin B<sub>12</sub> in the form of cyanocobalamin (Cat. No. 1152009), with a purity of 1.04% (10.4  
89 µg/mg), was bought from US Pharmacopeial Convention (USP, USA) to be used as the  
90 reference standard. The standard material (100 mg) was dissolved in water in a 100 mL  
91 volumetric flask to prepare a 10 mg/L stock solution. This stock solution was serially diluted  
92 with water to prepare 25, 50, 100, 250, and 500 µg/L working solutions.

### 94 **Development of sample preparation**

95 A previously reported sample preparation method (Kirchner et al., 2012; Moon et al., 2018)  
96 was used to remove protein, fat, and starch from the sample, after a slight modification of the  
97 method. Five grams of the cereal infant formula sample was placed on a 55 mL screw cap tube  
98 and dissolved in 49 mL of 0.2 M sodium acetate. The pH of the sample solution was adjusted  
99 to 4.0 to remove casein, which comprises ~80% of the milk protein fraction. Lowering the pH  
100 beyond 4.0 (isoelectric point of casein) resulted in isoelectric precipitation. Following this, 0.5

101 mL of 1% sodium cyanide was added and mixed, and the sample was extracted ultrasonically  
102 at 25°C for 10 min. After the addition of 0.5 mL of  $\alpha$ -amylase, the sample was incubated for  
103 30 min at 40°C and then for 30 min at 100°C in an incubator to initiate the reaction. Next, 20  
104 mL of the above solution was filtered by a Whatman paper and transferred to an immunoaffinity  
105 column (Easi-Ex-tract Vitamin B<sub>12</sub>, r-Biopharm, Glasgow, UK). The column was washed with  
106 10 mL water and injected with 40 mL of air by syringe to dry it. The loaded sample was eluted  
107 with 3 mL of methanol. The eluate was volatilized to dryness and then reconstituted in 0.5 mL  
108 of water. This was used as the test sample.

109

#### 110 **Chromatography parameters**

111 Chromatographic conditions were determined based on previously reported analogous  
112 methods that used LC–UV. A Shimadzu HPLC system (Shimadzu, Japan) equipped with a  
113 Shiseido Capcell Pak C18 UG 120 column (4.6 mm x 250 nm, 5  $\mu$ m) was used for the analysis  
114 of vitamin B<sub>12</sub>. Water and acetonitrile were used as the mobile phases for gradient elution. A  
115 flow rate of 1.0 mL/min and a column temperature of 35°C were maintained, and the injection  
116 volume was 50  $\mu$ L. HPLC grade solvents were filtered through a 0.45  $\mu$ m membrane and  
117 ultrasonically degassed prior to use. The specific chromatography conditions are A (water): B  
118 (acetonitrile) gradient system 0 - 3.4 min (100:0), 3.5 - 10.9 min (75:25), 11.0 - 18.9 (65:35),  
119 19 - 20 min (90:10), and 20 - 26 min (100:0).

120

#### 121 **Method validation**

122 Selectivity for vitamin B<sub>12</sub> detection was determined by comparing the chromatographic  
123 peaks of the test sample with those of the standard solutions. Linearity was assessed by  
124 injecting 25 to 500  $\mu$ g/L of vitamin B<sub>12</sub> solutions in duplicate. Qualitative parameters were

125 determined by comparing the retention times of the standard solution with those of the samples.  
126 The analyte was quantified from the calibration plot equations calculated by the least-squares  
127 method. Precision was calculated in terms of intra-day ( $n = 3$ ) and inter-day repeatabilities ( $n$   
128  $= 3$ ) by analyzing spiked cereal infant formula samples and was evaluated by calculating the  
129 relative standard deviation (RSD). Accuracy of the method was determined by calculating the  
130 recovery and appropriate standard deviation (SD) in cereal infant formula samples spiked with  
131 different amounts of vitamin B<sub>12</sub>. Detection limits were assessed in terms of limit of detection  
132 (LOD, signal-to-noise ratio (S/N) = 3) and limit of quantitation (LOQ, S/N = 10).

133

## 134 **Results and Discussion**

### 135 **Development of pretreatment method**

136 Analysis using the current method proposed by the MFDS is complex; moreover, it does not  
137 yield a desirable peak resolution in the analysis of powdered milk samples. In addition, the  
138 MFDS has not yet provided an appropriate method for the analysis of powder milk containing  
139 starch. Although the reason for the low peak resolution is not clear, the unstable nature of starch,  
140 proteins, and fats during sample treatment has been assumed to be a limitation in this  
141 conventional method. In addition, it is difficult to detect vitamin B<sub>12</sub> in some food samples  
142 using only one pretreatment method  
143 ([http://foodsafetykorea.go.kr/foodcode/01\\_03.jsp?idx=324](http://foodsafetykorea.go.kr/foodcode/01_03.jsp?idx=324)). Since vitamin B<sub>12</sub> exists in  
144 different forms at very low concentrations in powdered milk containing cereal, the sample  
145 preparation methodology is extremely crucial (Lee et al., 2015). In this study, individual  
146 pretreatment methods were developed by modifying the Association of Official Analytical  
147 Chemists (AOAC) method to detect vitamin B<sub>12</sub> in powdered milk containing starch in high  
148 amounts. In the modified AOAC method, samples were purified using an immunoaffinity



149 column and then subjected to HPLC to quantitate vitamin B<sub>12</sub> in the samples. Sodium cyanide  
150 and  $\alpha$ -amylase were used to remove starch, as mentioned in the experimental section. The pre-  
151 treatment involving clean-up and concentration using an immunoaffinity column enabled the  
152 efficient separation of trace amounts of vitamin B<sub>12</sub> from powdered milk samples. As a result  
153 of the sample pretreatment, vitamin B<sub>12</sub> was eluted at 9.3 min in the HPLC run, suggesting its  
154 efficient separation from the degradation products. This method allowed the separation and  
155 detection of vitamin B<sub>12</sub> within 10 min (Fig. 1). Detection using this approach under the  
156 described experimental conditions was slightly more rapid compared to that under the  
157 experimental conditions employed in a previous study (Heudi et al., 2006).

158

#### 159 **Method validation**

160 The specificity of the proposed technique was ensured by employing the well-established  
161 method of using highly selective immunoaffinity column for sample preparation (Nakos et al.,  
162 2017; Anatol et al., 2019). Detection limit and powdered milk containing starch recovery were  
163 determined by quantitative analysis, and certified reference material, SRM 1849a, was used to  
164 validate our analytical HPLC method. The amount of vitamin B<sub>12</sub> recovered in the SRM 1849a  
165 reference was 54.10  $\mu\text{g}/\text{kg}$ . Compared with value of 48.20  $\mu\text{g}/\text{kg}$  (given SRM 1843a certified  
166 value), the test represented recovery of the authentication value of 112.24%. The external  
167 calibration curve of vitamin B<sub>12</sub> standard solutions was linear in the range of 25–500  $\mu\text{g}/\text{L}$ , with  
168  $r^2 > 0.9999$ . The equation of the calibration curve was  $y = 53.806x - 150.44$ , where y represents  
169 the peak area of the curve obtained through UV detection, and x is the concentration ( $\mu\text{g}/\text{L}$ ) of  
170 vitamin B<sub>12</sub>. It is evident that the correlation coefficients were greater than 0.9999, which  
171 indicated a good correlation between the concentration and peak area of the investigated  
172 compounds. Accuracy was assessed by adding a known amount of the analyte, followed by

173 calculating the recovery using standards. Accuracy of the method was satisfactory, ranging  
174 from 83.41% to 106.57%, which was well within the recovery range reported for other food  
175 matrices (Zironi et al., 2013; Chamlagain et al., 2015). Intra-day and inter day variations were  
176 used to determine the precision of the established method. As shown table 1, RSD of intra-day  
177 and inter-day variations for compound was less than 5.31% and 3.90, respectively. The LOD  
178 and LOQ were 2.71 and 8.21  $\mu\text{g/L}$ , respectively (Table 2). These results suggest that the HPLC  
179 method involving sample pretreatment, immunoaffinity column separation is precise, accurate  
180 and sensitive for quantitative determination of active compounds in powdered milk containing  
181 starch.

182

### 183 **Monitoring test cereal infant formulas**

184 Four different powdered milk containing starch samples, of which two were manufactured  
185 in South Korea and two were manufactured in USA, were analyzed using the method developed  
186 in this study. The sample pre-treatment was repeated three times for each sample; the results  
187 are presented in Table 2. It is evident from Table 2 that the vitamin B<sub>12</sub> content in powdered  
188 milk products was in the range of 11.03 – 42.18  $\mu\text{g/kg}$ . As determined from the HPLC analysis,  
189 all the products contained trace nutrients that were higher than those displayed on the content  
190 labels. Therefore, the vitamin B<sub>12</sub> content displayed in powdered milk packaging available in  
191 the South Korean markets was well verified.

192

193

## 194 **Conclusion**

195 The nutrition labeling system of foods is being strengthened to provide appropriate  
196 information to consumers while choosing a food product. Therefore, there is an increasing need  
197 for scientifically established analytical techniques to strengthen the national management of  
198 foods with high nutritional components. In this work, sample pretreatment, immunoaffinity  
199 column separation, and HPLC were employed in combination to develop an analytical method  
200 for the extraction of vitamin B<sub>12</sub>. In the proposed method, starch was removed using a small  
201 quantity of  $\alpha$ -amylase, unlike the traditional methods. The validation results indicated high  
202 sensitivity and good accuracy and precision. The recovery and relative standard deviations  
203 were in the acceptable range. Additionally, the value obtained for the certified reference  
204 material (SRM 1849a) was within the range of certificated values. The developed method based  
205 on HPLC and sample pretreatment for the detection of vitamin B<sub>12</sub> could reduce the analysis  
206 time and manual labor, thereby proving to be an appropriate alternative to conventional  
207 analytical methods. Although, there are several methods for the detection of vitamin B<sub>12</sub> in  
208 dairy products, powdered milk etc., this is the first study to attempt the rapid detection of  
209 vitamin B<sub>12</sub> in powdered milk containing starch. Moreover, a beginner can be expected to easily  
210 perform this analytical procedure because of its simplicity. This method for the analysis of  
211 vitamin B<sub>12</sub> may be utilized in industries for micronutrient analysis in dairy products, functional  
212 foods, as well as powdered milk.

## 213 **Conflicts of Interest**

214 The authors declare no potential conflict of interest.

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219

### 220 **Author Contributions**

221 Conceptualization was Jin Man Kim, investigation, experiment and writing was Jung Min  
222 Park, and review & editing was Jong Ho Koh and Jin Man Kim.

223

### 224 **Ethics approval**

225 This paper does not require IRB/IACUC approval because there are no human and animal  
226 participants.

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295

296 **Table 1. Liquid chromatography (LC) conditions for vitamin B<sub>12</sub>**

Parameter	Condition			
Column	UG 120 C18 4.6 X 250nm, 5 μm, Shimadzu			
Detector	UV 361 nm			
Mobile phase	A: water	Time (min)	Solvent (A) %	Solvent (B) %
	B: Acetonitrile	0	100	0
	Gradient system	3.5	75	25
		11.0	65	35
		19.0	90	10
		20.0	100	0
		26.0	100	0
Flow rate	1.0 mL/min			
Column temperature	35°C			
Run time	25 min			
Injection volume	50 μL			

297



298 **Table 2. Inter-day and inter-day precision of vitamin B<sub>12</sub>**

299 All values are mean SD of three replicates.

Precision	Recovery (%)	SD	RSD <sup>1)</sup> (%)
Intra-day precision	103.72	5.51	5.31
	98.96	5.08	5.13
	93.61	3.94	4.21
	84.69	0.83	0.98
Inter-day precision	106.57	2.53	2.37
	95.02	2.05	2.16
	89.59	3.49	3.90
	83.41	2.01	2.40

300 <sup>1)</sup> RSD, relative standard deviation.

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302 **Table 3. Validation factors and monitoring test for vitamin B<sub>12</sub> in certified reference material**  
 303 **(SRM 1849a)**

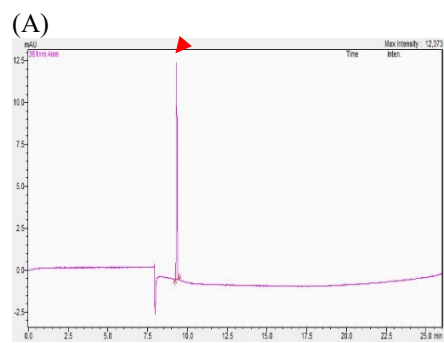
	Tested value (µg/kg)	RSD <sup>1)</sup> (%)	Recovery (%)
SRM 1849a	54.10±0.84	1.88	112.24±2.11
Samples	Tested value (µg/kg)		
Cereal infant formula	T-1	T-2	T-3
	11.93±2.08	11.03±0.16	42.18±1.57
			T-4
			16.65±1.18
r <sup>2</sup>	0.999		Linear Regression y= 53.806x-150.44
LOD <sup>2)</sup>	2.71 µg/L		Range 25-500 µg/L
LOQ <sup>3)</sup>	8.21 µg/L		

304 All values are mean SD of three replicates.

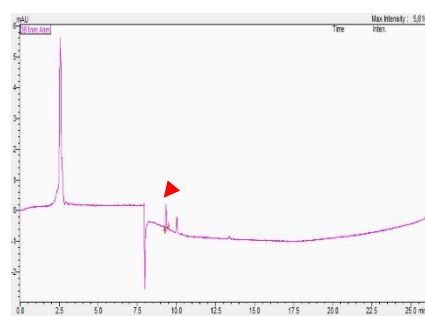
305 <sup>1)</sup> RSD, relative standard deviation; <sup>2)</sup> LOD, limit of detection; <sup>3)</sup> LOD, limit of quantitation.

306

307



(B)



308

309 **Fig. 1. Chromatogram of vitamin B<sub>12</sub>, (A) Standard of vitamin B<sub>12</sub>, (B) Powdered milk containing**  
310 **starch**

311

312

ACCEPTED